

Review

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Central adrenal insufficiency: open issues regarding diagnosis and glucocorticoid treatment

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Abstract

Background: Central adrenal insufficiency (CAI) is characterized by impaired adrenocorticotropin (ACTH) secretion because of a disease or injury to the hypothalamus or the pituitary, leading to a reduced cortisol production. CAI suspicion arises more frequently in patients with pituitary tumors, cranial irradiation/surgery/injury/infections, as well as after exogenous glucocorticoid withdrawal. Nevertheless, a late diagnosis is not uncommon because CAI may present with nonspecific signs or symptoms, as fatigue or hyponatremia.

Content: The PubMed database was searched (years 1980–2018), using “central adrenal insufficiency” and “ACTH deficiency” as keywords. Subsequently, reference sections of the retrieved articles were searched.

Summary: Dynamic tests are needed when morning basal cortisol levels are not sufficient to exclude or to confirm CAI. Short Synacthen Test (SST) is the most used, and Endocrine Society’s guidelines recommend a cortisol peak >500 nmol/L to exclude CAI. Despite thresholds, understanding the pretest probability of ACTH deficiency (the clinical background of the patient) is essential because the diagnostic accuracy of SST in case of a negative result is suboptimal. Glucocorticoid replacement therapy, able to replicate cortisol circadian rhythm, is required in patients with CAI; fludrocortisone treatment is not necessary. Short-acting glucocorticoid drugs (hydrocortisone or cortisone acetate) are the most used; lower doses than previously used are nowadays recommended

to reduce cortisol-related comorbidities. Promising results have been obtained with modified-release hydrocortisone, especially regarding glucose metabolism in patients with primary adrenal insufficiency.

Outlook: An accurate clinical diagnosis and a careful individualized therapy are mandatory in patients with CAI.

Keywords: adrenal insufficiency; central hypoadrenalism; corticotropin test; hydrocortisone.

Introduction

A functional hypothalamic-pituitary-adrenal (HPA) axis is essential for normal health and life expectancy. Adrenal insufficiency is defined by the inability of the adrenal cortex to produce a sufficient amounts of glucocorticoids (GC) and/or mineralocorticoids [1]. Patients with primary adrenal insufficiency (PAI) present with symptoms that result from the lack of both GC and mineralocorticoid secretion [2]; however, central adrenal insufficiency (CAI) is characterized by inappropriate adrenocorticotropin (ACTH) secretion as a result of disease or injury to the hypothalamus or pituitary gland, leading to a failure of adrenal cortisol production [3, 4].

Most signs and symptoms of adrenal insufficiency are nonspecific and present chronically as fatigue, weight loss, postural dizziness and hypotension; however, an acute presentation (adrenal crisis) with severe hypotension, abdominal pain (mimicking acute abdomen) and hyponatremia is a life-threatening condition [1, 2, 5, 6]. Mineralocorticoid secretion is preserved in patients with CAI; therefore, hyperkalemia and hypotension are typical features in PAI, as well as hyperpigmentation due to enhanced secretion of pro-opiomelanocortin peptides [1, 2]. Late diagnosis is not uncommon; several authors reported that CAI seems to be a frequently overlooked cause of hospitalization in case of hyponatremia, especially in the elderly [7, 8].

Prompt and correct diagnosis is mandatory because adequate hormonal replacement therapy is lifesaving

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[3, 4]. The measurement of morning serum cortisol is the initial step of diagnosis; however, when cortisol levels are not clearly sufficient or insufficient to respectively exclude or confirm adrenal insufficiency, biochemical testing is recommended. Different basal or stimulated cortisol cutoff levels, mainly developed with different immunoassays, have been proposed in literature; however, in clinical practice, it is mandatory to consider the pretest probability of HPA axis impairment (in terms of the clinical background of the patient) [3, 4, 9, 10].

Patients with adrenal insufficiency require lifelong GC replacement therapy, whose primary aim is to provide an appropriate amount of cortisol at the appropriate time, closely to the circadian cortisol rhythm [3, 11–13]. One of the major drawbacks related to GC substitution therapy is the lack of objective methods or biomarkers able to reveal under- or overreplacement. Therefore, the treatment is guided both by physician's expertise and by subjective health status of the individual [3, 4, 13].

This review focuses on the diagnosis and management of CAI, considering assay issues, cortisol stimulation and the titration of substitutive treatment.

Is secondary adrenal insufficiency a rare condition? Who should be tested?

Impaired ACTH secretion could be related to endogenous hypothalamic-pituitary injury or to iatrogenic HPA axis suppression by synthetic GC. In Table 1, we summarize the high-risk population for CAI.

A GC treatment is largely used in general population (up to 2%): CAI onset after GC discontinuation is not uncommon and sometimes unrecognized (up to 4% of patients after nasal GC) [14]. There is no administration form, dosing, treatment duration or underlying disease that could exclude CAI onset, although higher doses and longer use of GC give the highest risk [14, 15]. Considering hospitalized patients with critical illness, transient HPA axis suppression is increasingly observed and reported as relative adrenal insufficiency, a combination of primary and secondary adrenal insufficiency, because both low CRH-ACTH levels and impaired adrenal gland steroidogenesis are observed [16]. Another form of combined primary

Table 1: Etiology and prevalence of acquired CAI.

Sellar mass
Craniopharyngioma (87%)
Pituitary adenoma (secreting and non functioning, up to 40% pre and 75% postsurgery)
Pituitary carcinoma or metastases
Other skull base tumors
Drug induced
Withdrawal of exogenous GCs (from 7% for asthma with inhalation GC to 60% for hematological malignancies)
Surgery for Cushing's syndrome (up to 100% of patients in remission, recovery of HPA function in 3–24 months in most patients)
Immune checkpoint inhibitors (ipilimumab, a form of iatrogenic hypophysitis, up to 20% of treated patients)
Post intracranial procedures
Pituitary or intracranial surgery (up to 50%)
Pituitary irradiation (ranging from 12% to 68%, prevalence increases with time)
Cranial or total-body irradiation for non-pituitary tumors (up to 10%)
Infiltrative
Neurosarcoidosis (up to 49%)
Histiocytosis (up to 10%)
Hemochromatosis (up to 45%)
Inflammatory
Hypophysitis (up to 60%)
Meningitis (particularly tuberculous)
Traumatic/vascular
Pituitary apoplexy
Traumatic brain injury (up to 8%)
Subarachnoid hemorrhage (up to 6%)
Miscellaneous
Idiopathic
Empty sella syndrome (up to 15%)
Relative adrenal insufficiency in hospitalized patients with acute illness (from 10% in hospitalized ill patient to 60% in those with septic shock)
Liver cirrhosis (all stages, 10%–82% of cirrhotics depending on the test used)

and secondary adrenal insufficiency may occur in liver cirrhosis, where adrenal cortisol secretion is impaired, because of low levels of total cholesterol, and ACTH secretion is reduced, secondary to increased circulating proinflammatory cytokines [17].

Because hypoadrenalism after surgical treatment of endogenous Cushing's syndrome is one of the remission criteria, the development of CAI starts early after the resection of an ACTH- or a cortisol-secreting tumor. Therefore, in this setting, GC substitutive treatment must be started early after surgery and tapered upon physiological recovery of HPA axis [4, 18].

Besides the aforementioned conditions, endogenous CAI is a rare condition, and its prevalence is about 150–280 cases per million inhabitants [4], higher than that reported for PAI (100–140 cases per million) [1]. In patients with pituitary tumors, cranial surgery or irradiation history or brain injury infection, CAI is identified in the formal screening process for hypopituitarism. About one-third of the patients with pituitary failure may present CAI [3, 4, 10, 16, 19–22]. Considering treatment options for pituitary adenoma, medical therapy seems to be protective regarding the integrity of HPA axis, especially in secreting forms [23]. The reported prevalence of CAI after trans-sphenoidal surgery could be up to 50% in patients with pituitary adenoma [24–28], and 90% for craniopharyngioma, which usually requires larger or repeated surgical approaches, often combined with radiotherapy [29, 30]. Despite CAI diagnosis, patients need to be reassessed in the follow-up because the HPA axis could recover after surgery in some cases, avoiding unnecessary GC treatment [24]. In 2015, Yedinak et al. [26] reported that the complete recovery rate of HPA axis after surgery in patients with acromegaly was higher than those with non-functioning pituitary adenoma, despite the need for medical therapy, after 7 years of follow-up using 1 µg Short Synacthen Test (SST). They discovered 20 new onset of CAI, developed 6–12 weeks after pituitary surgery in a cohort of 100 patients, especially in macroadenomas: 70% of these new onset CAI resolved after 1 year of observation [26].

CAI is common after pituitary irradiation (ranging from 12% to 68% [22, 31, 32]), as well as after cranial radiation for intracerebral or nasopharyngeal tumors and total body irradiation for hematological malignancies. Despite initial cancer, total radiation dose or kind of treatment (being fractionated, proton beam or stereotactic radiotherapy), CAI may take a few years to develop, with incidence of all pituitary hormone deficiencies almost doubling between years 2 and 7 of follow-up [33, 34].

Selected high-risk categories of patients are those after brain injury (traumatic or subarachnoid hemorrhage [13]),

infectious meningitis or pituitary abscess [19, 35], and hypophysitis induced by ipilimumab, a monoclonal antibody directed against cytotoxic T-lymphocyte antigen-4 acting as an immune checkpoint inhibitor [21, 36].

Reversible, acquired and isolated ACTH deficiency is particularly associated with lymphocytic hypophysitis: the association with pregnancy, the presence of antipituitary antibodies and the radiological features (homogeneous pituitary expansion with stalk thickened) are the clinical hallmarks of hypophysitis [37, 38].

Another critical issue common in hypopituitarism concerns the GH replacement therapy, which could unmask a latent CAI: GH is able to accelerate the peripheral metabolism of cortisol, revealing an unknown CAI or requiring an increase in current GC dose [39].

What is the rationale for SST?

The measurement of morning serum cortisol is the initial step of CAI diagnosis: different cutoffs have been proposed; however, it is generally accepted that low morning cortisol levels are indicative of adrenal insufficiency, in both primary and secondary forms. The threshold of morning cortisol level that exclude adrenal insufficiency with a sensitivity of 100% is uncertain: the choice of a higher cutoff allows to consider also partial HPA axis impairment (common in CAI). The final diagnosis of adrenal insufficiency is highly likely if morning cortisol is <138 nmol/L (5 µg/dL, two samples if clinically feasible) in PAI [1, 2], as well as in CAI [40, 41]. However, to avoid false diagnosis and inappropriate GC treatment, stricter criteria (<80–100 nmol/L) in one sample are suggested to diagnose CAI [3, 4, 10].

One of the major drawbacks concerning the diagnosis of adrenal insufficiency (and unfortunately one of the less known) is that the reliability of routine immunoassays is highly variable because an accurate cortisol quantification is compromised by matrix effects, considerable interassay variation and partial antibody specificity, therefore leading to an uncertainty area starting in the range of 100–150 nmol/L [42]. Lack of standardization across different platforms for cortisol measurement is an issue in routine clinical practice because some cutoff values established with immunoassay could not be used with other platforms [43]. Liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS) offers improved specificity and sensitivity; however, cortisol cutoffs proposed in guidelines did not consider cortisol assays [1, 10, 44].

Elevated levels of cortisol-binding globulin in women receiving estrogen treatment may increase total serum cortisol levels; therefore, in some cases, a woman with suspected CAI may present a misleading normal serum cortisol concentration [43]. Contrariwise, morning salivary cortisol level is free: it has been studied in women using oral contraceptive pill or during pregnancy [45], and in suspected adrenal insufficiency with immunometric assay [46] or with LC-MS/MS [47]. However, the diagnostic accuracy of salivary cortisol in suspected adrenal insufficiency is questionable [48], and its routine use is not yet suggested [1, 10]. High ACTH values and cutaneous hyperpigmentation increase the clinical suspicion of PAI [1, 2]; nonetheless, the finding of a random low/normal ACTH level may not be helpful in diagnosing CAI [3, 4].

When cortisol levels are not clearly sufficient or insufficient to respectively exclude or confirm adrenal insufficiency, biochemical testing with SST is recommended to confirm the diagnosis [10]. Despite the guideline's recommendation, there are a large amount of data showing that almost all patients with an early morning cortisol >400 nmol/L pass stimulation tests if modern assays for cortisol are considered, i.e. the recent automated immunoassays Advia Centaur (Siemens), Architect (Abbott) or the Roche Modular System (Roche) [4, 49].

The rationale of SST in CAI is the assumption that the acute responsiveness of the adrenal zona fasciculata is reduced in chronic endogenous corticotropin deficiency; therefore, a rapid ACTH injection fails to stimulate a cortisol response after 30–60 min [40]. However, it is to consider that a patient with partial ACTH deficiency (i.e. with a suboptimal cortisol peak after SST) is likely to have ACTH levels within normal values for most of the day, without functional adrenal atrophy, thus explaining the relative low sensitivity of SST in some cases with partial HPA axis impairment.

Standard (or high, 250 μg) dose SST is recommended in patients with PAI [1]; however, in CAI, both low (1 μg) and standard dose SSTs are suggested: a cortisol peak >500 nmol/L at 30 or 60 min is sufficient to exclude CAI [10]. In clinical practice, a significant deviation from this threshold is adopted in 52% of the 250- μg SST and 39% of the 1- μg SST, reflecting the use of locally derived cutoffs, dependent on the assay platform used and the clinical context of the patient [49, 50]. In this scenario, understanding the pretest probability of ACTH deficiency (the clinical background of the patient: a positive history for pituitary surgery or irradiation, previous or concomitant GC treatment, exposure to cranial or total body irradiation and so on) is essential to properly diagnose CAI because SST had a suboptimal likelihood ratio for a negative test

[9]. To avoid analytical interferences, Endocrine Society's guidelines suggest testing for HPA axis at least 18–24 h after the last hydrocortisone (HC) dose, or after a longer withdrawal for long-acting GC [10].

Because the rationale for SST is the assumption that cortisol response is blunted in chronic ACTH deficiency, it is important to define when we consider the condition that probably will induce the CAI as “chronic”. Few papers investigated the optimal time shift to perform SST (suggesting that it can be carried out at least 4 weeks after pituitary surgery [51, 52]), but the major part of the studies proposed the stimulation test 2–3 months after surgery [53–56]. Therefore, in clinical practice, SST is scheduled 2–3 months after every HPA axis injury. At the best of our knowledge, conclusive data regarding the time of cortisol testing in other different etiologies of CAI (i.e. withdrawal of exogenous GCs, brain injury and so on) are lacking: further studies are needed to establish the optimal time shift to assess HPA axis in these patients.

Standard or low-dose SST: that is the question

There is an ongoing debate about the optimal dose of corticotropin in SST: it is suggested that 250 μg is a supraphysiological stimulus, eliciting very high circulating ACTH levels, therefore excessive to detect mild CAI [56]. In patients with partial endogenous ACTH deficiency, the adrenal responsiveness to high doses of ACTH may be preserved, with a resultant false-negative SST. By contrast, the 1- μg dose is known to be the lowest one to induce adrenal response because 0.6 or 0.8 μg was not able to increase cortisol levels in healthy controls [57].

Contrasting data about SST diagnostic accuracy are reported: some authors reported a superior role of 1 μg [58, 59] or 250 μg [52, 54, 60] SSTs, and others reported that both tests presented similar results [55, 56, 61].

Kazlauskaitė et al. [40] performed a systematic quantitative meta-analysis (600 patients) to compare the performance of the 250- or 1- μg SST in patients with CAI [confirmed with insulin tolerance test (ITT) or metyrapone test], considering only high-risk populations (pituitary or other intracranial tumors, brain surgery or irradiation and hypopituitarism). Half of the patients performed both tests: the 1- μg SST had a larger area under the receiver operating characteristic curve than the standard dose, also after adjusting for cortisol assay. Therefore, they concluded that 30-min cortisol values after the 1- μg SST presented a better diagnostic performance. Authors also

suggested a three-step approach for evaluating patients with possible CAI: the first was measuring morning cortisol levels, the second was to perform a 1- μg SST if serum cortisol levels were <138 or >365 nmol/L and the third was to consider ITT or metyrapone test only in indeterminate SST results. This flowchart would be able to diagnose the majority of patients; however, clinical judgment (starting from the clinical background of the patient) would maintain a critical role.

Recently, Ospina et al. [9] included only studies reporting CAI diagnosed with ITT or metyrapone test (1200 adult patients) in a meta-analysis. They concluded that both high and low-dose SSTs had low sensitivity (64% for standard and 83% for low-dose SST) and high specificity (93% for 250 μg and 86% for the 1- μg SST) to diagnose CAI, resulting in a reasonable positive likelihood ratio (9 and 6, respectively) but a suboptimal negative likelihood ratio (0.4 and 0.2, respectively). Therefore, SST is more helpful in ruling in the condition when positive (to confirm CAI in patients with inadequate cortisol response to ACTH), rather than to rule out when negative (to exclude CAI in patients with negative tests, considered as normal cortisol response to ACTH) [9]. Therefore, also the second meta-analysis confirmed that it is mandatory to consider the pretest probability of CAI before prescribing an SST.

To conclude, there is no general consensus regarding which SST is superior because contrasting data are available [52, 54–56, 58–61], the results of the first and second meta-analyses are discordant [9, 40] and finally the recent guidelines do not suggest which SST to use (1 or 250 μg) [10]. In our clinical practice, we use the 1- μg SST (to reduce false-negative results) combined with the clinical history of the patient (to rule out CAI in case of suboptimal cortisol response).

How should SST be prepared and performed?

During the 250- μg SST, cortisol levels are obtained at baseline (in the morning between 8:00 and 9:00 a.m.) and after ACTH injection (most commonly at 30 or 60 min). Some authors found that 30 min is the maximum response for the low dose, and then cortisol levels started to decrease, whereas using the high-dose SST, there is still a little cortisol increase from 30 to 60 min, not only in healthy controls but also in patients with known impaired HPA axis [55].

The concentration of 250 μg is a ready-to-use formulation (Synacthen[®] or Cortrosyn[®]); otherwise, 1 $\mu\text{g}/\text{mL}$ is obtained by diluting a vial of ACTH (available in 250 $\mu\text{g}/\text{mL}$)

in 249 mL of sterile saline physiological solution NaCl 0.9% and then aspirating 1 mL of solution [23]. Another preparation is to extract 0.4 mL from a 1-mL vial of 250 μg and add it to a 100-mL physiological solution, resulting in an ACTH concentration of 100 $\mu\text{g}/100$ mL, and then 1 mL of this solution could be injected [56]. Some authors suggest to add all the 250- μg ampule to 49 mL of 0.9% saline, then adding 0.2 mL of these latter solution (5 $\mu\text{g}/\text{mL}$) to 0.8 mL of 0.9% saline (obtaining 1 $\mu\text{g}/\text{mL}$) [62]. To conclude, up to 10 dilution methods for the 1- μg SST have been reported, resulting in dose variation (up to 0.8 μg) and so in false results, with potentially important clinical sequelae [50].

Another interesting, and crucial if unknown, issue of SST is the loss of ACTH through tubing. In 2010, Wade et al. [63] measured the ACTH concentrations in both diluted and administered solutions, reporting that about 25 cm of tubing was able to reduce the amount of injected ACTH of 22–59%.

To conclude, loss of ACTH during dilution (with different protocol adopted in different centers) or administration might sharply reduce the cortisol response when 1 μg of ACTH is used. In this scenario, a 1 μg vial is needed from the factory, and direct venous injection is suggested to reduce, at least in part, some bias connected to corticotropin test.

How to manage indeterminate/unreliable SST?

Because CAI is a life-threatening condition, both false-negative and false-positive SST results are detrimental for the patients. Other tests have been used, or are currently used, as ITT, metyrapone test or glucagon test (summarized in Table 2). Although the CRH test has a theoretical role to distinguish hypothalamic from pituitary disease, it is no longer recommended [3, 10].

ITT is considered the gold standard to diagnose CAI because the response of the HPA axis to hypoglycemia (<2.2 mmol/L) constitutes an evidence of sufficient physiological stress to provoke ACTH release, not only in case of surgical stress [3, 4, 10]. ITT could be used also to assess the integrity of somatotroph function; however, other tests are suggested to diagnose GH deficiency [64, 65]. As SST, it is unnecessary to perform an ITT in subjects with low morning cortisol levels. Insulin and glucagon tests could be performed in acute setting, early after suspected HPA axis injury, when the knowledge of the necessity for GC replacement therapy is paramount. Despite the

Table 2: Pros and cons of screening tests used to diagnose CAI.

Test	Pros	Cons	Interpretation of the results
Morning (unstimulated) serum cortisol	<ul style="list-style-type: none"> – Easily available – Included in Endocrine Society's guidelines 	<ul style="list-style-type: none"> – Assay dependent (reduced accuracy in case of low cortisol levels) – Requires withdrawal of synthetic GCs 	<ul style="list-style-type: none"> – Screening test for PAI and CAI – Conclusive diagnosis if low cortisol levels (with different cutoffs, generally accepted <1.38 nmol/L) or high cortisol levels (with different cutoffs, generally accepted >500 nmol/L)
Short Synacthen Test (SST)	<ul style="list-style-type: none"> – Easy to perform – Well tolerated and safe – Good sensitivity (adequate to rule in CAI); high likelihood of having ACTH deficiency in patients that do not respond – Recommended in the Endocrine Society's guidelines 	<ul style="list-style-type: none"> – Unreliable if recent onset of CAI – Do not assess somatotroph function – Poor specificity (inadequate to rule out CAI when negative) – Assay dependent 	<ul style="list-style-type: none"> – Confirmation test for PAI and CAI – 250-μg SST in PAI, 1 μg or 250-μg SST in CAI – 250-μg SST: peak cortisol <500 nmol/L at 30 or 60 min indicate PAI or CAI – 1-μg SST: peak cortisol <400–500 nmol/L at 30 min indicate CAI
Insulin tolerance test	<ul style="list-style-type: none"> – High diagnostic accuracy – Assessment of corticotroph and somatotroph function – Could be performed early after pituitary injury 	<ul style="list-style-type: none"> – Labor intensive and time consuming, unpleasant for patients – Contraindicated in ischemic heart and brain disease – Risk of false-negative results if significant hypoglycemia is not achieved 	<ul style="list-style-type: none"> – Used to assess hypothalamic-pituitary dysfunction in CAI – 0.05–0.15 U/kg insulin – Cortisol peak >500–550 nmol/L to exclude CAI
Metyrapone test	<ul style="list-style-type: none"> – Easy to perform 	<ul style="list-style-type: none"> – Not included in the Endocrine Society's guidelines – 11-deoxycortisol is not measured worldwide – Lack of standardization 	<ul style="list-style-type: none"> – Confirmation test for CAI – 30 mg/kg metyrapone – ACTH >17 pmol/L and/or 11-deoxycortisol >200 nmol/L to exclude CAI
Glucagon test	<ul style="list-style-type: none"> – Assessment of corticotroph and somatotroph function – Performed when ITT is contraindicated 	<ul style="list-style-type: none"> – Risk of adrenal insufficiency in patients with CAI – Not included in the Endocrine Society's guidelines – Nausea and vomiting in up to 30% cases – Suboptimal cortisol response in 8% of healthy individuals – Time consuming – Lack of standardization – Not included in the Endocrine Society's guidelines 	<ul style="list-style-type: none"> – Used to assess hypothalamic-pituitary dysfunction in CAI – 1 mg glucagon – cortisol peak >500–550 nmol/L to exclude CAI

risks correlated to hypoglycemia, the main limitations of the ITT are that it requires the supervision of physicians or nurses and it is contraindicated in the elderly or in those individuals with seizures or cardiocerebrovascular disease.

Overnight metyrapone test is considered convenient and sensitive, compared with the ITT; the diagnostic accuracy of this test is related to an increase in 11-deoxycortisol levels >200–260 nmol/L [55, 62]. Metyrapone is administered orally 30 mg/kg before sleeping, and blood is sampled the following morning for 11-deoxycortisol and ACTH. When directly compared with SST, some authors reported that metyrapone test presented an increased [62] or similar [55] sensitivity to diagnose CAI. Nevertheless, metyrapone is not available worldwide (i.e. in the US), and metyrapone test is not mentioned in the Endocrine Society's guidelines for hypopituitarism [10]. Further studies are needed to assess its role in the diagnostic flow-chart of CAI.

GC replacement therapy in CAI

A careful cost–benefit analysis is suggested in the era of personalized medicine because a combination of various treatment (especially surgery and radiotherapy) could increase the risk of hypopituitarism and CAI-related mortality [22, 23, 66–69].

Patients with CAI require lifelong GC replacement therapy, whose primary aim is to replicate as much as possible the circadian cortisol rhythm [3, 11–13]. Substitutive therapy in adrenal insufficiency should be able to provide an appropriate amount of cortisol at the appropriate time: one of the major drawbacks related to currently available oral GC formulations is their lack of ability to properly replicate the physiologic cortisol circadian rhythm. Therefore, well-being is often not fully restored in all patients, and life expectancy may even be reduced [12, 70].

Recently, it has been established that the daily cortisol production rate ranges between 5 and 10 mg/m² of body surface area [71], which is lower than that previously considered. Traditionally, the daily HC dose was 30 mg/day, split into two or three doses (with the highest in the morning); given the recent discovery of lower levels of cortisol production rates, the “traditional 30 mg of HC” was supraphysiological [70]. Moreover, higher doses of HC replacement therapy (>25 mg/day) are related to increased mortality in patients with acromegaly, especially for cardiovascular disease (up to 44% mortality if HC therapy was >30 mg/day) [69]. Also, patients with

non-functioning pituitary adenoma that developed CAI presented increased mortality if receiving >20 mg/day of HC [28]. All synthetic compounds could be used to treat adrenal insufficiency, but in clinical practice, short half-life GCs [HC and cortisone acetate (CA)] are preferred than those with longer half-life (as prednisolone, usually given as a single morning dose of 3–5 mg [72], or dexamethasone 0.375–0.5 mg, preferred in adults with congenital adrenal hyperplasia [73]).

Filipsson et al. [74] have described, in a large series of more than 1500 patients with ACTH deficiency, an adverse metabolic profile in patients taking higher doses of GCs, with increased total cholesterol, triglycerides, waist circumference and glycosylated hemoglobin compared with the ACTH-sufficient patients. Intriguingly, those subjects who had HC-equivalent doses <20 mg/day did not differ in metabolic end points compared with the ACTH-sufficient patients. Moreover, they observed that patients receiving HC were at high risk to develop an adverse metabolic profile (especially regarding glucose metabolism) than those using CA.

After the synthesis of cortisone in 1950 [70], the treatment regimens remained almost unchanged: the short elimination half-life of HC (approximately 1.5 h) when given in traditional immediate-release preparations twice or thrice daily leads, however, to high peaks with low trough values in between. More than 1200 patients with adrenal insufficiency (11% with CAI) reported in a web-based survey that there is the need for improvement in the daily management of adrenal insufficiency because the negative impact of their treatment on subjective health was consistent [75].

Recently, novel HC preparations (dual-release HC) have been developed, allowing a cortisol serum profile closer to circadian secretion, improving the treatment of patients with CAI [76–79]. In PAI, dual-release HC is able to ameliorate glucose metabolism, to improve cardiovascular risk factors (especially weight, waist circumference, blood pressure and cholesterol levels) [76, 78] and to reduce recurrent infections [80]. On the contrary, there really are too few data in CAI to recommend the use of dual-release HC to improve metabolic parameters.

Management of therapy adequacy

In clinical practice, achieving the correct replacement dose is a challenge, especially to identify the lowest GC dose that relieves symptoms of insufficiency, while avoiding cortisol excess.

Several authors proposed serum, salivary or urinary biomarkers, but none has been extensively studied. Mah et al. [81] suggested a normative graph to evaluate the replacement therapy. Considering the nomogram, one sample of serum cortisol 2.5–5 h after the first administration of GC therapy compared with reference percentiles should be used for individual adjustment. Nevertheless, this protocol requires blood sample collection and could be inconvenient to manage for both outpatients and physicians; moreover, it has been used only in the original 20 patients of the cohort and not replicated by other groups.

In 2012, we described that multiple saliva collections could be used to measure salivary cortisol and to compute area under the curve (to assess daily GC exposure), in a small group of patients with CAI treated with CA. Multiple saliva sampling, a convenient easy-to-manage and stress-free tool for outpatients, could pave the way to a future application of salivary cortisol assays for assessing the adequacy of replacement therapy because samples could be performed before and after GC dose [47, 82]. Recently, we reported, in a proof-of-concept prospective study, that the reduction of GC treatment (assessed with daily salivary cortisol) resulted in a better replication of circadian cortisol rhythm and in an improvement of diastolic blood pressure [83].

Urinary free cortisol excretion fails to detect daily fluctuations of cortisol (secondary to treatment) and suffers of a marked day-to-day variability depending on renal function [11, 84]. A clinical score was proposed by Arlt et al. [85], but subsequent studies did not support the superiority of this qualitative system over quantitative methods.

Controversies and areas of uncertainty

The long-term natural history of patients with pituitary disease and the risk to develop CAI during the follow-up are still unknown, as well as reliable biomarkers to assess the adequacy of substitutive GC therapy. Therefore, efforts should be considered to answer the following questions:

- The choice between standard (250 µg) or low (1 µg) SST in CAI is still uncertain, despite several case-control studies and two meta-analyses.
- How to manage patients with low/normal cortisol levels and inadequate response to SST. Obviously, the proposed thresholds are based on the meta-analyses

of well-designed studies and considering ITT as gold standard for AI diagnosis; nevertheless, for those patients with an intermediate morning serum cortisol measurement (200 nmol/L) and suboptimal peak after SST (i.e. 350 nmol/L) or for whom observation is not appropriate, other diagnostic tools should be performed.

- The impact in mimicking the physiological circadian cortisol profile by dual-release HC formulations has to be further investigated in patients with CAI, as well as the close relationship among cortisol substitutive therapy and other treatments (in order of replacement levothyroxine, estrogens and growth hormone).
- How to assess the adequacy of cortisol treatment is a critical issue because several clinical or biochemical markers have been proposed and none has been shown to be superior in routine clinical practice.

Conclusions

A functional HPA axis is essential for normal health and life expectancy. Therefore, CAI is a life-threatening disorder associated with increased morbidity and mortality: correct diagnosis and treatment are crucial for well-being and lifesaving.

The milestones of CAI are not only the measurements of basal serum cortisol and dynamic tests: the medical history is crucial because the diagnostic accuracy biochemical of tests is not definitely reliable in all patients. Nevertheless, SST is one of the most used, at least 6–8 weeks after surgery, because cortisol response is blunted in chronic ACTH deficiency.

The management of therapy adequacy needs a careful evaluation because both excessive and insufficient treatments are correlated to increased morbidity and mortality.

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References

1. Bornstein SR, Allolio B, Arlt W, Barthel A, Don-Wauchope A, Hammer GD, et al. Diagnosis and treatment of primary adrenal insufficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2016;101:364–89.
2. Husebye ES, Allolio B, Arlt W. Consensus statement on the diagnosis, treatment and follow-up of patients with primary adrenal insufficiency. *J Intern Med* 2014;275:104–15.
3. Grossman AB. The diagnosis and management of central hypoadrenalism. *J Clin Endocrinol Metab* 2010;95:4855–63.
4. Crowley RK, Argese N, Tomlinson JW, Stewart PM. Central hypoadrenalism. *J Clin Endocrinol Metab* 2014;99:4027–36.
5. Hahner S, Loeffler M, Bleicken B, Drechsler C, Milovanovic D, Fassnacht M, et al. Epidemiology of adrenal crisis in chronic adrenal insufficiency: the need for new prevention strategies. *Eur J Endocrinol* 2010;162:597–602.
6. Hahner S, Spinnler C, Fassnacht M, Burger-Stritt S, Lang K, Milovanovic D, et al. High incidence of adrenal crisis in educated patients with chronic adrenal insufficiency: a prospective study. *J Clin Endocrinol Metab* 2015;100:407–16.
7. Chanson P. Severe hyponatremia as a frequent revealing sign of hypopituitarism after 60 years of age. *Eur J Endocrinol* 2003;149:177–8.
8. Diederich S, Franzen NF, Bahr V, Oelkers W. Severe hyponatremia due to hypopituitarism with adrenal insufficiency: report on 28 cases. *Eur J Endocrinol* 2003;148:609–17.
9. Ospina NS, Al Nofal A, Bancos I, Javed A, Benkhadra K, Kapoor E, et al. ACTH stimulation tests for the diagnosis of adrenal insufficiency: systematic review and meta-analysis. *J Clin Endocrinol Metab* 2016;101:427–34.
10. Fleseriu M, Hashim IA, Karavitaki N, Melmed S, Murad MH, Salvatori R, et al. Hormonal replacement in hypopituitarism in adults: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2016;101:3888–921.
11. Debono M, Price JN, Ross RJ. Novel strategies for hydrocortisone replacement. *Best Pr Res Clin Endocrinol Metab* 2009;23:221–32.
12. Debono M, Ross RJ, Newell-Price J. Inadequacies of glucocorticoid replacement and improvements by physiological circadian therapy. *Eur J Endocrinol* 2009;160:719–29.
13. Grossman A, Johannsson G, Quinkler M, Zelissen P. Therapy of endocrine disease: perspectives on the management of adrenal insufficiency: clinical insights from across Europe. *Eur J Endocrinol* 2013;169:R165.
14. Broersen LH, Pereira AM, Jørgensen JO, Dekkers OM. Adrenal insufficiency in corticosteroids use: systematic review and meta-analysis. *J Clin Endocrinol Metab* 2015;100:2171–80.
15. Dinsen S, Baslund B, Klose M, Rasmussen AK, Friis-Hansen L, Hilsted L, et al. Why glucocorticoid withdrawal may sometimes be as dangerous as the treatment itself. *Eur J Intern Med* 2013;24:714–20.
16. Reddy P. Clinical approach to adrenal insufficiency in hospitalised patients. *Int J Clin Pract* 2011;65:1059–66.
17. Karagiannis AK, Nakouti T, Pipili C, Cholongitas E. Adrenal insufficiency in patients with decompensated cirrhosis. *World J Hepatol* 2015;7:1112.
18. Nieman LK, Biller BM, Findling JW, Murad MH, Newell-Price J, Savage MO, et al. Treatment of Cushing's syndrome: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2015;100:2807–31.
19. Tsiakalos A, Xynos ID, Sipsas NV, Kaltsas G. Pituitary insufficiency after infectious meningitis: a prospective study. *J Clin Endocrinol Metab* 2010;95:3277–81.
20. Aimaretti G, Ambrosio MR, Di Somma C, Gasperi M, Cannavò S, Scaroni C, et al. Residual pituitary function after brain injury-induced hypopituitarism: a prospective 12-month study. *J Clin Endocrinol Metab* 2005;90:6085–92.
21. Brilli L, Danielli R, Ciuoli C. Prevalence of hypophysitis in a cohort of patients with metastatic melanoma and prostate cancer treated with ipilimumab. *Endocrine* 2017;58:535–41.
22. Darzy KH. Radiation-induced hypopituitarism. *Curr Opin Endocrinol Diabetes Obes* 2013;20:342–53.
23. Ceccato F, Lizzul L, Zilio M, Barbot M, Denaro L, Emanuelli E, et al. Medical treatment for acromegaly does not increase the risk of central adrenal insufficiency: a long-term follow-up study. *Horm Metab Res* 2016;48:514–9.
24. Webb SM, Rigla M, Wägner A, Oliver B, Bartumeus F. Recovery of hypopituitarism after neurosurgical treatment of pituitary adenomas. *J Clin Endocrinol Metab* 1999;84:3696–700.
25. Jahangiri A, Wagner J, Han SW, Tran MT, Miller LM, Tom MW, et al. Rate and time course of improvement in endocrine function after more than 1000 pituitary operations. *Neurosurgery* 2014;61(Suppl 1):163–6.
26. Yedinak C, Hameed N, Gassner M, Brzana J, McCartney S, Fleseriu M. Recovery rate of adrenal function after surgery in patients with acromegaly is higher than in those with non-functioning pituitary tumors: a large single center study. *Pituitary* 2015;18:701–9.
27. Berker M, Hazer DB, Yücel T, Gürlek A, Cila A, Aldur M, et al. Complications of endoscopic surgery of the pituitary adenomas: analysis of 570 patients and review of the literature. *Pituitary* 2012;15:288–300.
28. Hammarstrand C, Ragnarsson O, Hallén T, Andersson E, Skoglund T, Nilsson AG, et al. Higher glucocorticoid replacement doses are associated with increased mortality in patients with pituitary adenoma. *Eur J Endocrinol* 2017;177:251–6.
29. Crowley RK, Hamnvik OP, O'Sullivan EP, Behan LA, Smith D, Agha A, et al. Morbidity and mortality in patients with craniopharyngioma after surgery. *Clin Endocrinol (Oxf)* 2010;73:516.
30. Wijnen M, van den Heuvel-Eibrink M, Janssen J. Very long-term sequelae of craniopharyngioma. *Eur J Endocrinol* 2017;176:755–67.
31. Minniti G, Traish D, Ashley S, Gonsalves A, Brada M. Fractionated stereotactic conformal radiotherapy for secreting and nonsecreting pituitary adenomas. *Clin Endocrinol (Oxf)* 2006;64:542–8.
32. Li X, Li Y, Cao Y, Li P, Liang B, Sun J, et al. Safety and efficacy of fractionated stereotactic radiotherapy and stereotactic radiosurgery for treatment of pituitary adenomas: a systematic review and meta-analysis. *J Neurol Sci* 2017;372:110–16.
33. Appelman-Dijkstra NM, Kokshoorn NE, Dekkers OM, Neelis KJ, Biermasz NR, Romijn JA, et al. Pituitary dysfunction in adult patients after cranial radiotherapy: systematic review and meta-analysis. *J Clin Endocrinol Metab* 2011;96:2330–40.
34. Kyriakakis N, Lynch J, Orme SM, Gerrard G, Hatfield P, Loughrey C, et al. Pituitary dysfunction following cranial radiotherapy for adult-onset nonpituitary brain tumours. *Clin Endocrinol (Oxf)* 2016;84:372–9.
35. Gao L, Guo X, Tian R, Wang Q, Feng M, Bao X, et al. Pituitary abscess: clinical manifestations, diagnosis and treatment of 66 cases from a large pituitary center over 23 years. *Pituitary* 2017;20:189–94.

36. Torino F, Corsello SM, Salvatori R. Endocrinological side-effects of immune checkpoint inhibitors. *Curr Opin Oncol* 2016;28:278–87.
37. Lupi I, Manetti L, Raffaelli V, Lombardi M, Cosottini M, Iannelli A, et al. Diagnosis and treatment of autoimmune hypophysitis: a short review. *J Endocrinol Invest* 2011;34:e245.
38. Glezer A, Bronstein M. Pituitary autoimmune disease: nuances in clinical presentation. *Endocrine* 2012;42:74–9.
39. Scaroni C, Ceccato F, Rizzati S, Mantero F. Concomitant therapies (glucocorticoids and sex hormones) in adult patients with growth hormone deficiency. *J Endocrinol Invest* 2008;31(Suppl):61–5.
40. Kazlauskaitė R, Evans AT, Villabona CV, Abdu TA, Ambrosi B, Atkinson AB, et al. Corticotropin tests for hypothalamic-pituitary-adrenal insufficiency: a metaanalysis. *J Clin Endocrinol Metab* 2008;93:4245–53.
41. Chanson P, Guignat L, Goichot B, Chabre O, Boustani DS, Reynaud R, et al. Adrenal insufficiency: screening methods and confirmation of diagnosis. *Ann Endocrinol (Paris)* 2017;78:495–511.
42. Hawley JM, Owen LJ, Lockhart SJ, Monaghan PJ, Armston A, Chadwick CA, et al. Serum cortisol: an up-to-date assessment of routine assay performance. *Clin Chem* 2016;62:1220–9.
43. Gounden V, Rampursat YD, Jialal I. Secretory tumors of the pituitary gland: a clinical biochemistry perspective. *Clin Chem Lab Med* 2018;57:150–64.
44. El-Farhan N, Rees DA, Evans C. Measuring cortisol in serum, urine and saliva—are our assays good enough? *Ann Clin Biochem* 2017;54:308–22.
45. Manetti L, Rossi G, Grasso L, Raffaelli V, Scattina I, Del Sarto S, et al. Usefulness of salivary cortisol in the diagnosis of hypercortisolism: comparison with serum and urinary cortisol. *Eur J Endocrinol* 2013;168:315–21.
46. Ceccato F, Barbot M, Zilio M, Ferasin S, Occhi G, Daniele A, et al. Performance of salivary cortisol in the diagnosis of Cushing's syndrome, adrenal incidentaloma, and adrenal insufficiency. *Eur J Endocrinol* 2013;169:31–6.
47. Ceccato F, Selmin E, Sabbadin C, Dalla Costa M, Antonelli G, Plebani M, et al. Improved salivary cortisol rhythm with dual-release hydrocortisone. *Endocr Connect* 2018;7:965–97.
48. Raff H. Utility of salivary cortisol measurements in Cushing's syndrome and adrenal insufficiency. *J Clin Endocrinol Metab* 2009;94:3647–55.
49. Sbardella E, Isidori AM, Woods CP, Argese N, Tomlinson JW, Shine B, et al. Baseline morning cortisol level as a predictor of pituitary-adrenal reserve: a comparison across three assays. *Clin Endocrinol (Oxf)* 2017;86:177–84.
50. Cross AS, Helen Kemp E, White A, Walker L, Meredith S, Sachdev P, et al. International survey on high- and low-dose synacthen test and assessment of accuracy in preparing low-dose synacthen. *Clin Endocrinol (Oxf)* 2018;88:744–51.
51. Dökmetaş HS, Colak R, Keleştimur F, Selçuklu A, Unlühizarci K, Bayram F. A comparison between the 1-microg adrenocorticotropin (ACTH) test, the short ACTH (250 microg) test, and the insulin tolerance test in the assessment of hypothalamic-pituitary-adrenal axis immediately after pituitary surgery. *J Clin Endocrinol Metab* 2000;85:3713–9.
52. Courtney CH, McAllister AS, Bell PM, McCance DR, Leslie H, Sheridan B, et al. Low- and standard-dose corticotropin and insulin hypoglycemia testing in the assessment of hypothalamic-pituitary-adrenal function after pituitary surgery. *J Clin Endocrinol Metab* 2004;89:1712–7.
53. Ambrosi B, Barbetta L, Re T, Passini E, Faglia G. The one microgram adrenocorticotropin test in the assessment of hypothalamic-pituitary-adrenal function. *Eur J Endocrinol* 1998;139:575–9.
54. Tordjman K, Jaffe A, Trostanetsky Y, Greenman Y, Limor R, Stern N. Low-dose (1 µg) adrenocorticotrophin (ACTH) stimulation as a screening test for impaired hypothalamo-pituitary-adrenal axis function: sensitivity, specificity and accuracy in comparison with the high-dose (250 µg) test. *Clin Endocrinol (Oxf)* 2000;52:633–40.
55. Giordano R, Picu A, Bonelli L, Balbo M, Berardelli R, Marinazzo E, et al. Hypothalamus-pituitary-adrenal axis evaluation in patients with hypothalamo-pituitary disorders: comparison of different provocative tests. *Clin Endocrinol (Oxf)* 2008;68:935–941.
56. Dekkers OM, Smit JW, Romijn JA, Pereira AM. Comparison of the cortisol responses to testing with two doses of ACTH in patients with suspected adrenal insufficiency. *Eur J Endocrinol* 2011;164:83–7.
57. Dickstein G, Spigel D, Arad E, Shechner C. One microgram is the lowest ACTH dose to cause a maximal cortisol response. There is no diurnal variation of cortisol response to submaximal ACTH stimulation. *Eur J Endocrinol* 1997;137:172–5.
58. Abdu TA, Elhadd TA, Neary R, Clayton RN. Comparison of the low dose short synacthen test (1 µg), the conventional dose short synacthen test (250 µg), and the insulin tolerance test for assessment of the hypothalamo-pituitary-adrenal axis in patients with pituitary disease. *J Clin Endocrinol Metab* 1999;84:838–43.
59. Maghnie M, Uga E, Temporini F, Di Iorgi N, Secco A, Tinelli C, et al. Evaluation of adrenal function in patients with growth hormone deficiency and hypothalamic-pituitary disorders: comparison between insulin-induced hypoglycemia, low-dose ACTH, standard ACTH and CRH stimulation tests. *Eur J Endocrinol* 2005;152:735–41.
60. Cho HY, Kim JH, Kim SW, Shin CS, Park KS, Kim SW, et al. Different cut-off values of the insulin tolerance test, the high-dose short Synacthen test (250 µg) and the low-dose short Synacthen test (1 µg) in assessing central adrenal insufficiency. *Clin Endocrinol* 2014;81:77–84.
61. Mayenknecht J, Diederich S, Bähr V, Plöckinger U, Oelkers W. Comparison of low and high dose corticotropin stimulation tests in patients with pituitary disease. *J Clin Endocrinol Metab* 1998;83:1558–62.
62. Soule S, Van Zyl Smit C, Parolis G, Attenborough S, Peter D, Kinvig S, et al. The low dose ACTH stimulation test is less sensitive than the overnight metyrapone test for the diagnosis of secondary hypoadrenalism. *Clin Endocrinol (Oxf)* 2000;53:221–7.
63. Wade M, Baid S, Calis K, Raff H, Sinaii N, Nieman L. Technical details influence the diagnostic accuracy of the 1 microg ACTH stimulation test. *Eur J Endocrinol* 2010;162:109–13.
64. Corneli G, Di Somma C, Baldelli R, Rovere S, Gasco V, Croce CG, et al. The cut-off limits of the GH response to GH-releasing hormone-arginine test related to body mass index. *Eur J Endocrinol* 2005;153:257–64.
65. Molitch ME, Clemmons DR, Malozowski S, Merriam GR, Vance ML. Evaluation and treatment of adult growth hormone deficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2011;96:1587–609.
66. Schöfl C, Grussendorf M, Honegger J, Tönjes A, Thyroke-Gronostay D, Mayr B, et al. Failure to achieve disease control

- in acromegaly: cause analysis by a registry-based survey. *Eur J Endocrinol* 2015;172:351–6.
67. Puig Domingo M. Treatment of acromegaly in the era of personalized and predictive medicine. *Clin Endocrinol (Oxf)* 2015;83:3–14.
 68. Newell-Price J, Trainer P, Besser M, Grossman A. The diagnosis and differential diagnosis of Cushing's syndrome and pseudo-Cushing's states. *Endocr Rev* 1998;19:647–72.
 69. Sherlock M, Reulen RC, Alonso AA, Ayuk J, Clayton RN, Sheppard MC, et al. ACTH deficiency, higher doses of hydrocortisone replacement, and radiotherapy are independent predictors of mortality in patients with acromegaly. *J Clin Endocrinol Metab* 2009;94:4216–23.
 70. Hahner S, Allolio B. Therapeutic management of adrenal insufficiency. *Best Pract Res Clin Endocrinol Metab* 2009;23:167–79.
 71. Kraan GP. The daily cortisol production reinvestigated in healthy men. The serum and urinary cortisol production rates are not significantly different. *J Clin Endocrinol Metab* 1998;83:1247–52.
 72. Smith DJ, Prabhudev H, Choudhury S, Meeran K. Prednisolone has the same cardiovascular risk profile as hydrocortisone in glucocorticoid replacement. *Endocr Connect* 2017;6:766–72.
 73. Merke DP. Approach to the adult with congenital adrenal hyperplasia due to 21-hydroxylase deficiency. *J Clin Endocrinol Metab* 2008;93:653–60.
 74. Filipsson H, Monson JP, Koltowska-Hägström M, Mattsson A, Johannsson G. The impact of glucocorticoid replacement regimens on metabolic outcome and comorbidity in hypopituitary patients. *J Clin Endocrinol Metab* 2006;91:3954–61.
 75. Forss M, Batcheller G, Skrtic S, Johannsson G. Current practice of glucocorticoid replacement therapy and patient-perceived health outcomes in adrenal insufficiency—a worldwide patient survey. *BMC Endocr Disord* 2012;12:8.
 76. Johannsson G, Nilsson AG, Bergthorsdottir R, Burman P, Dahlqvist P, Ekman B, et al. Improved cortisol exposure-time profile and outcome in patients with adrenal insufficiency: a prospective randomized trial of a novel hydrocortisone dual-release formulation. *J Clin Endocrinol Metab* 2012;97:473–81.
 77. Quinkler M, Miodini Nilsen R, Zopf K, Venz M, Øksnes M. Modified-release hydrocortisone decreases BMI and HbA_{1c} in patients with primary and secondary adrenal insufficiency. *Eur J Endocrinol* 2015;172:619.
 78. Giordano R, Guaraldi F, Marinazzo E, Fumarola F, Rampino A, Berardelli R, et al. Improvement of anthropometric and metabolic parameters, and quality of life following treatment with dual-release hydrocortisone in patients with Addison's disease. *Endocrine* 2016;51:360–8.
 79. Nilsson AG, Bergthorsdottir R, Burman P. Long-term safety of once-daily, dual-release hydrocortisone in patients with adrenal insufficiency: a phase 3b, open-label, extension study. *Eur J Endocrinol* 2017:360–8.
 80. Isidori AM, Venneri MA, Graziadio C, Simeoli C, Fiore D, Hasenmajer V, et al. Effect of once-daily, modified-release hydrocortisone versus standard glucocorticoid therapy on metabolism and innate immunity in patients with adrenal insufficiency (DREAM): a single-blind, randomised controlled trial. *Lancet Diabetes Endocrinol* 2018;6:173–85.
 81. Mah PM, Jenkins RC, Rostami-Hodjegan A, Newell-Price J, Doane A, Ibbotson V, et al. Weight-related dosing, timing and monitoring hydrocortisone replacement therapy in patients with adrenal insufficiency. *Clin Endocrinol (Oxf)* 2004;61:367–75.
 82. Ceccato F, Albiger N, Reimondo G, Frigo AC, Ferasin S, Occhi G, et al. Assessment of glucocorticoid therapy with salivary cortisol in secondary adrenal insufficiency. *Eur J Endocrinol* 2012;167:769–76.
 83. Ceccato F, Barbot M, Lizzul L, Selmin E, Saller A, Albiger N, et al. Decrease in salivary cortisol levels after glucocorticoid dose reduction in patients with adrenal insufficiency: a prospective proof-of-concept study. *Clin Endocrinol (Oxf)* 2018;88:201–8.
 84. Monson JP. The assessment of glucocorticoid replacement therapy. *Clin Endocrinol (Oxf)* 1997;46:269–70.
 85. Arlt W, Rosenthal C, Hahner S, Allolio B. Quality of glucocorticoid replacement in adrenal insufficiency: clinical assessment vs. timed serum cortisol measurements. *Clin Endocrinol (Oxf)* 2006;64:384–9.