Diclofenac is one of the most widely prescribed non-steroidal anti-inflammatory drugs (NSAIDs). Use of diclofenac is associated with two major limitations; first, rare, but serious and sometimes fatal, gastrointestinal (GI) side-effects, including ulceration, and hemorrhage, especially in the elderly (1, 2), and second, poor water solubility.

Pharmacosomes are amphiphilic lipid vesicular systems that have shown their potential in improving the bioavailability of poorly water soluble as well as poorly lipophilic drugs. Diclofenac is a poorly water soluble drug and also causes gastrointestinal toxicity. To improve the water solubility of diclofenac, its pharmacosomes (phospholipid complex) have been prepared and evaluated for physicochemical analysis. Diclofenac was complexed with phosphatidylcholine (80 %) in equimolar ratio, in the presence of dichloromethane, by the conventional solvent evaporation technique. Pharmacosomes thus prepared were evaluated for drug solubility, drug content, surface morphology (by scanning electron microscopy), phase transition behaviour (by differential scanning calorimetry), crystallinity (by X-ray powder diffraction) and in vitro dissolution. Pharmacosomes of diclofenac were found to be irregular or disc shaped with rough surfaces in SEM. Drug content was found to be 96.2 ± 1.1 %. DSC thermograms and XRPD data confirmed the formation of the phospholipid complex. Water solubility of the prepared complex was found to be 22.1 µg mL⁻¹ as compared to 10.5 µg mL⁻¹ of diclofenac. This improvement in water solubility in prepared pharmacosomes may result in improved dissolution and lower gastrointestinal toxicity. Pharmacosomes showed 87.8 % while the free diclofenac acid showed a total of only 60.4 % drug release at the end of 10 h of dissolution study.

Keywords: diclofenac, solubility, pharmacosomes, phospholipid complex

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Different approaches have been applied to decrease NSAID-induced GI toxicity. For example, association of NSAIDs with phospholipids has been suggested to improve GI safety of these drugs (3). The presence of an adsorbed layer of surface-active phospholipids on the surface of the mucus that covers the surface epithelium is suggested to protect the GI tissues by providing a hydrophobic layer between the epithelium and the luminal contents (4, 5). It has been reported that NSAIDs associated with zwitterionic phospholipids may reduce GI toxicity (3).

Besides GI toxicity of diclofenac, it is also poorly water soluble, due to which its dissolution in GI fluid is very low, which in turn adversely affects the bioavailability (50–60 % only). The association of diclofenac with zwitterionic phospholipids, which may be both electrostatic and hydrophobic in nature, renders the phospholipids more water-soluble and the NSAID more lipid-soluble (6). It has been reported that the diffusion of NSAIDs across lipid membranes and into target cells is accelerated when they are in a complex with phosphatidylcholine (PC) (3).

Therefore developing drugs as lipid complexes (pharmacosomes) may be a potential approach to improve solubility and to minimize the GI toxicity of diclofenac. Pharmacosomes are amphiphilic drug-lipid complexes, which are stable and more bioavailable drug delivery systems with low interfacial tension between the system and the GI fluid, thereby facilitating the membrane, tissue, or cell wall transfer in the organism (7).

This work aims to develop and characterize the pharmacosomes of diclofenac along with its in vitro drug release study.

**EXPERIMENTAL**

**Materials**

Diclofenac potassium was obtained as a gift sample from Wings Pharma, India. Soya phosphatidylcholine (LIPOID S-80) was obtained as a gift sample from LIPOID, Germany. All other chemicals were of analytical grade.

**Formulation of pharmacosomes**

Diclofenac salt was converted into the acid form to provide an active hydrogen site for complexation. Diclofenac acid was prepared by acidification of an aqueous solution of diclofenac potassium, extraction into chloroform, and subsequent recrystallization. Diclofenac-PC complex was prepared by associating diclofenac acid with an equimolar concentration of PC. The equimolar concentration of PC and diclofenac acid were placed in a 100-mL round bottom flask and dissolved in dichloromethane. The solvent was evaporated under vacuum at 40 °C in a rotary vacuum evaporator (Perfit Model No. 5600, Büchi type, Perfit Ltd., India). The pharmacosomes were collected as the dried residue and placed in a vacuum desiccator overnight and then subjected to characterization.
Drug content

To determine the drug content in pharmacosomes of diclofenac (diclofenac-PC complex), a complex equivalent to 50 mg diclofenac was weighed and added into a volumetric flask with 100 mL of pH 6.8 phosphate buffer. Then the volumetric flask was stirred continuously for 24 h on a magnetic stirrer. At the end of 24 h, suitable dilutions were made and measured for the drug content at 276 nm UV spectrophotometrically (double beam UV-Visible spectrophotometer, Lambda 25, Perkin Elmer, USA).

Solubility

To determine the change in solubility due to complexation, solubility of diclofenac acid and diclofenac-PC complex was determined in pH 6.8 phosphate buffer and n-octanol by the shake-flask method. Diclofenac acid (50 mg) (and 50 mg equivalent in case of complex) was placed in a 100-mL conical flask. Phosphate buffer pH 6.8 (50 mL) was added and then stirred for 15 minutes. The suspension was then transferred to a 250 mL separating funnel with 50 mL n-octanol and was shaken well for 30 minutes. Then the separating funnel was kept still for about 30 minutes. Concentration of the drug was determined from the aqueous layer spectrophotometrically at 276 nm.

Scanning electron microscopy (SEM)

To detect the surface morphology of the pharmacosomes, SEM of the complex was recorded on a scanning electron microscope (JEOL JSM 5600, Japan).

Differential scanning calorimetry (DSC)

Thermograms of diclofenac acid, phosphatidylcholine (80 %) and the diclofenac-PC complex were recorded using a 2910 Modulated Differential Scanning Calorimeter V4.4E (TA Instruments, USA). The thermal behavior was studied by heating 2.0 ± 0.2 mg of each individual sample in a covered sample pan under nitrogen gas flow. The investigations were carried out over the temperature range 25–250 °C at a heating rate of 10 °C min–1.

X-ray powder diffraction (XRPD)

The crystalline state of diclofenac in the different samples was evaluated using X-ray powder diffraction. Diffraction patterns were obtained on a Bruker Axs- D8 Discover Powder X-ray diffractometer, Germany. The X-ray generator was operated at 40 kV tube voltages and 40 mA tube current, using the Kα lines of copper as the radiation source. The scanning angle ranged from 1 to 60° of 2θ in the step scan mode (step width 0.4° min–1). Diclofenac acid, phosphatidylcholine 80 % (Lipoid S-80) and the prepared complex were analyzed. The results are shown in Fig. 3.
Dissolution study

In vitro dissolution studies of diclofenac complex as well as plain diclofenac acid were performed in triplicate in a USP (8) six station dissolution test apparatus, type II (Veego Model No. 6 DR, India) at 100 rpm and at 37 °C. An accurately weighed amount of the complex equivalent to 100 mg of diclofenac acid was put into 900 mL of pH 6.8 phosphate buffer. Samples (3 mL each) of dissolution fluid were withdrawn at different intervals and replaced with an equal volume of fresh medium to maintain sink conditions. Withdrawn samples were filtered (through a 0.45-μm membrane filter), diluted suitably and then analyzed spectrophotometrically at 276 nm. Fig. 4 gives a graphical representation of the in vitro release profile from the formulations.

Statistical analysis

Results are expressed as mean values ± standard deviations and the significance of the difference observed was analyzed by Student’s t-test.

RESULTS AND DISCUSSION

Pharmacosomes of diclofenac were prepared with an equimolar ratio (1:1) of diclofenac and phosphatidylcholine in the presence of dichloromethane by the conventional solvent evaporation technique. The content of diclofenac in the pharmacosome, as estimated by UV spectrophotometry, was found to be 96.2 ± 1.1% (m/m). Pharmacosomes showed a high percentage of drug loading, which is specific to them. Complexation provides good percent loading of the drug, which makes the drug delivery clinically feasible. On the other hand, in liposomes of diclofenac sodium, the encapsulation efficiency was found to be just 59% (9). Special methods such as coating of these vesicular systems were needed to improve the loading of the drug. This is not required in the pharmacosomes in which the drug is reversibly bonded chemically with the lipids and thus shows not only good percent loading but also better stability than in liposomes.

Water solubility of diclofenac from pharmacosomes was found to be much higher than that of diclofenac acid. Table I provides the solubility data. The increase in solubility of diclofenac acid in the complex can be explained by the fact that solubilization resulted from the micelle formation in the medium and by the amorphous characteristics of the complex. As amphiphilic surfactants, phospholipids could increase the solubility.

<table>
<thead>
<tr>
<th>Drug</th>
<th>Solubility</th>
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<tbody>
<tr>
<td></td>
<td>Aqueous layer (μg mL⁻¹)ᵃ</td>
</tr>
<tr>
<td>Diclofenac acid</td>
<td>10.5 ± 1.5</td>
</tr>
<tr>
<td>Diclofenac pharmacosomes</td>
<td>22.1 ± 2.4</td>
</tr>
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</table>

ᵃ Mean ± SD; n = 3.
of the drug by the action of wetting and dispersion (3, 6). Unlike the non-polar nature of diclofenac, the complex showed amphiphilic nature, which may prove to be responsible for improved bioavailability of the drug (10).

Scanning electron micrographs of the complex are shown in Fig. 1. Pharmacosomes were found to be irregular or disc shaped with rough surface morphology. The complex was found to involve free flowing particles. Phospholipids are natural components, so their different purity grades may have different effects on the shape and surface morphology (11).

In order to substantiate the association of diclofenac acid with PC, DSC analysis was performed on diclofenac acid, PC, and the pharmacosomes of diclofenac. Results confirmed the association of diclofenac acid and PC in the complex as both peaks representing diclofenac acid and PC changed positions. Phospholipids (Fig. 2b) showed two major peaks at 83.21 °C and 107.90 °C and a small peak at 64.45 °C. The first peak of phospholipids is a mild peak (at 64.45 °C), which is probably due to the hot movement of the phospholipid polar head group. The second (83.21 °C) peak is very sharp, which appears to be due to the phase transition from gel to liquid crystalline state. The non-polar hydrocarbon tail of phospholipids may be melted during this phase, yielding a sharp peak. This melting might have occurred in two phases that subsequently gave another peak (107.90 °C), which is relatively less sharp. Diclofenac acid (Fig. 2a) showed a sharp endothermic peak at 171.31 °C. On the other hand, pharmacosomes of diclofenac (Fig. 2c) showed a broad peak at 68.34 °C, which is different from the peaks of the individual components of the complex. It is evident that the original peaks of diclofenac and phospholipids disappear from the thermogram of pharmacosomes (complex) and the phase transition temperature is lower than that of phospholipids. The DSC thermograms of the phospholipid complexes of some phytoconstituents, like silybin, puerarin, curcumin and some xanthones, revealed similar results (12–14).

To check whether the changes in the diclofenac crystal morphology correspond to a polymorphic transition and to study the solid state of diclofenac phospholipid complex, XRPD analysis was conducted. The XRPD of diclofenac complex revealed a broad peak similar to PC (Fig. 3). It suggested that the diclofenac in the phospholipid complex was either in amorphous form or molecularly dispersed. These results are well supported by
our previous studies done with the phospholipid complexes of aspirin (11). The disapp-
pearance of diclofenac crystalline diffraction peaks confirmed the formation of the pho-
spholipid complex. Unlike liposomes, chemical bonding between the drug and phospho-
lipids in the development of pharmacosomes might have resulted into a significant change
of its X-ray diffraction.

Fig. 2. DSC thermograms of: a) diclofenac acid, b) phosphatidylcholine, and c) pharmacosome of
diclofenac.
The pharmacosomes of diclofenac showed a better dissolution profile than diclofenac acid (Fig. 4). Unlike free diclofenac acid (which showed only 60.4% drug release at the end of 10 h) the diclofenac complex showed 87.8% at the end of 10 h of the dissolution study in pH 6.8 phosphate buffer. Solid dissolution is a complex operation influ-

![Fig. 3. X-ray powder diffraction (XRPD) study of: a) diclofenac acid, b) phosphatidylcholine and c) pharmacosome of diclofenac.](image)

![Fig. 4. Dissolution study of pharmacosomes of diclofenac and diclofenac acid (mean ± SD; n = 3).](image)
enced by a number of factors, not only by particle size. Differences in the crystal habit, surface area, surface energies, particle size and wettability may all play a role in affecting the dissolution rate of powder (15). Phospholipids being amphiphilic surfactants, increased the solubility of the drug by the action of wetting and dispersion. That is why the dissolution profile of the complex was found to be improved. In some studies done with silybin and xanthones, the \textit{in vitro} drug release from the complexes was found to be pH dependent and with the increase of the medium pH the drug dissolution was increased (12).

\section*{CONCLUSIONS}

In the present study, a diclofenac-phospholipid complex (pharmacosomes) was prepared by a simple and reproducible method and evaluated for various physicochemical parameters. Physicochemical investigations showed that diclofenac formed a stoichiometric complex with phospholipid with improved solubility and dissolution profile. DSC and XRPD studies confirmed the formation of the complex. Thus it can be concluded that the pharmacosomes of diclofenac may be of potential use for improving dissolution and for reducing the gastrointestinal toxicity of the drug.

Pharmacosomes may be developed also for other NSAIDs with poor bioavailability and GI side effects. Moreover, the pharmacosomes of phytoconstituents (with poor water and/or lipid solubility) may also be developed for improving their aqueous solubility and lipophilicity and hence bioavailability.

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\section*{REFERENCES}


**SAŽETAK**

**Razvoj i fizikokemijsko vrednovanje farmakosoma diklofenaka**

AJAY SEMALTY, MONA SEMALTY, DEVENDRA SINGH i M. S. M. RAWAT

Farmakosomi su amfibilni lipidni vezikularni sustavi sa sposobnošću poboljšanja bioraspoloživosti lijekova slabo topljivih u vodi i organskim otapalima. U svrhu povećanja topljivosti diklofenaka (lijekovite tvari koja je slabo vodotopljiva, a uzrokuje i gastro-intestinalnu toksičnost) pripravljeni su i evaluirani njegovi farmakosomi (fosfolipidni kompleksi). Diklofenak je kompleksiran s fosfatidilkolinom (80 %) u ekvimolarnom omjeru, u prisutnosti diklorometana, konvencionalnom metodom evaporačije. Tako pripravljenim farmakosomima ispitivana je topljivost, sadržaj lijekovite tvari, morfološka površine (pomoću pretražne elektronske mikroskopije), ponašanje pri prijelazu faza (pomoću diferencijalne pretražne kalorimetrije), kristaliničnost (rendgenskom analizom praha) i in...
oslobađanje. Farmakosomi diklofenaka su nepravilnog oblika ili u obliku diska te imaju neravnu površinu u SEM-u. Sadržaj ljekovite tvari je 96,2 ± 1,1 %. DSC termogrami i XRPD podaci potvrdili su nastajanje fosfolipidnog kompleksa. Topljivost u vodi dobivenih kompleksa bila je 22,1 µg mL⁻¹, a topljivost samog diklofenaka 10,5 µg mL⁻¹. Postignuto poboljšanje topljivosti može imati za posljedicu povećano oslobađanje i manju gastrointestinalnu toksičnost. Tijekom 10 h iz farmakosoma se oslobodilo 87,8 %, a iz slobodnog diklofenaka samo 60,4 % ljekovite tvari.

Ključne riječi: diklofenak, topljivost, farmakosomi, fosfolipid kompleks

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