This brief review summarises recent observations which suggest a possible mechanism for organophosphate-induced delayed neuropathy (OPIDN). Neuropathy target esterase (NTE) has been shown to deacylate endoplasmic reticulum (ER) membrane phosphatidylcholine (PtdCho). Raised levels of PtdCho are present in the brains of swiss cheese/NTE mutant Drosophila together with abnormal membrane structures, axonal and dendritic degeneration and neural cell loss. Similar vacuolated pathology is found in the brains of mice with brain-specific deletion of the NTE gene and, in old age, these mice show clinical and histopathological features of neuropathy resembling those in wild-type mice chronically dosed with tri-ortho-cresylphosphate. It is suggested that OPIDN results from the loss of NTE’s phospholipase activity which in turn causes ER malfunction and perturbation of axonal transport and glial-axonal interactions.

KEY WORDS: brain, carbamate, endoplasmic reticulum, organophosphate, phospholipid

During the 20th century, thousands of people were paralysed by accidental ingestion of tri-ortho-cresylphosphate (TOCP). The same paralysis was present in rare cases of poisoning by some organophosphorus (OP) insecticides. OP-induced delayed neuropathy (OPIDN) has been studied experimentally in chickens, and features a latent period of at least one week before the onset of progressive hind limb ataxia and paralysis accompanied by distal degeneration of long nerves in the spinal cord and legs (1, 2). Neuropathic OPs inhibit the ability of brain homogenates to catalyse hydrolysis of an artificial ester substrate and this enzyme activity is called NTE (3, 4). However, the events linking NTE to axon degeneration were enigmatic until molecular cloning of NTE (5) allowed new experimental approaches.

The biochemical function of NTE

Studies in mammalian cell lines and yeast have shown that NTE is anchored to the cytoplasmic face of the endoplasmic reticulum (ER) where it catalyses the deacylation of phosphatidylcholine (PtdCho) to glycerophosphocholine (6, 7) (Figure 1). PtdCho comprises up to 50% of the phospholipid content of eukaryotic cell membranes and thus NTE plays a major role in membrane homeostasis. PtdCho accumulates in, and NTE activity is absent from, the brains of Swiss cheese mutant Drosophila and this is accompanied by membrane abnormalities, axonal/dendritic degeneration and brain vacuolation (8). Likewise, in mice with brain-specific NTE deletion, prominent vacuolation of the hippocampus and thalamus is evident between 2 to 5 months of age (9).
Species susceptibility to OPIDN

In large animals (humans, cattle, cats, chickens) a single dose of neuropathic OP causes ataxia and paralysis within 1 to 3 weeks. By contrast, rodents, and particularly mice, are refractory to OPIDN. A few swollen axons in the brain stem of mice were detected three weeks after a single dose of TOCP, but no clinical signs or axonal degeneration (10). However, dosing mice with TOCP daily for 9 months induced axonal degeneration and hind-limb paralysis (11). Finally, brain-specific NTE knock-out mice exhibit progressive axonal degeneration and hind-limb ataxia between 6 to 12 months of age (Read et al, unpublished).

Non-neuropathic inhibitors of NTE

Carbamates such as pyridostigmine have been used to protect humans and animals from imminent exposure to OP nerve agents. The basis for this protection is that carbamylation of acetylcholinesterase (AChE) prevents its reaction with the OP during a critical period before the available concentration of the latter falls to non-toxic levels; during this period, continuous reactivation and re-inhibition of AChE by the carbamate must permit sufficient activity to prevent toxic accumulation of acetylcholine (4). Similarly, pre-dosing chickens with certain carbamates can protect against induction of OPIDN by a subsequent dose of neuropathic OP; this protection lasts as long as the half-life of carbamylated NTE (about 12 h) (12).

Much of the enigma surrounding the pathogenesis of OPIDN stems from the finding that compounds such as phenylmethanesulfonyfluoride (PMSF) and certain organophosphinates can also protect against OPIDN, even though they themselves appear to inhibit NTE for several days. Furthermore, neuropathic OPs undergo an aging reaction in which one of the alkyl/aryl groups is rapidly expelled from organophosphorylated NTE, leaving a negatively-charged adduct. This process renders the enzyme far more resistant to reactivation. However, it has been argued that the generation of negatively-charged adduct, rather than inhibition of NTE’s catalytic activity, is responsible for initiating OPIDN (13, 14) and that aging may induce a toxic gain of function in NTE (15).

On the other hand, sequential dosing with large amounts of a carbamate followed by PMSF does induce (mild) neuropathy. From this, it has been suggested that aging is not essential for OPIDN and that NTE inhibitors may have varying degrees of neuropathic efficacy, analogous to full and partial receptor agonists (16).

Inactivation of NTE initiates OPIDN

OP inhibition of NTE will, at least temporarily, disrupt ER phospholipid homeostasis which may cause general malfunction of the ER. Since this organelle is the starting point for all secretory transport in eukaryotic cells, both axonal transport and interaction between the axon and supporting glial cells will be perturbed. Inhibition of retrograde axonal transport has been demonstrated in OP-dosed chickens before the onset of ataxia (17). Abnormal glial-axonal/neuronal interaction is the earliest detected pathological change in the brains of swiss cheese flies (18). Distal parts of long axons may be most sensitive to this perturbation, as they are the farthest from the sources of energy and materials supplied by the neuronal cell body (Fig 2). The fact

Figure 1 NTE deacylates endoplasmic reticulum-associated phosphatidylcholine. In this biosynthetic pathway, choline is converted to phosphocholine, then CDP-choline which is finally reacted with diacylglycerol, forming phosphatidylcholine (PtdCho) at the cytoplasmic face of the ER membrane. NTE is tethered in this location by a transmembrane segment and deacylates PtdCho to form soluble products, free fatty acids (FFA) and glycerophosphocholine (GroPCho). For more details see ref. (7).

Figure 2 Long axons are hypervulnerable to loss of NTE function

1. Transient disruption of phospholipid homeostasis and ER function in both neuron and glial cells.
2. Impairment of both axonal transport and glial-axon interaction
3. Distal axons are most sensitive to loss of support and begins to degenerate.
that genetic knock-out of NTE and chronic TOCP dosing induce a similar neuropathic syndrome in mice is compelling evidence that the OP is indeed acting by inactivating NTE rather than by inducing a toxic gain of function.

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REFERENCES

Sažetak

DEGENERACIJA AKSONA I ESTERAZA POVEZANA S NEUROPATSKIM DJELOVANJEM ORGANOFOSFORNIH SPOJEVA - PREGLED

Ovim se kratkim pregledom razmatraju nedavna opažanja koja upućuju na mogući mehanizam odgodene neuropatije uzrokovane organofosfataima (engl. organophosphate-induced delayed neuropathy, krat. OPIDN). Za esterazu povezanu s neuropatskim djelovanjem organofosfornih spojeva (engl. neuropathy target esterase, krat. NTE) dokazano je da deacilira fosfatidilkinolin (PtdCho) membrane endoplazmatskog retikuluma (ER). Povišene razine PtdCho prisutne su u mozgu swiss cheese/NTE mutanta mušice Drosophila uz abnormalne membranske strukture, degeneraciju aksona i dendrita te gubitak neurona. Slična je vakuolarna patologija zamijenjena u mozgu miševa u kojima je obrisan NTE gen u mozgu te koji u starijoj dobi pokazuju kliničke i histopatološke znakove neuropatije koja je slična onoj u običnim miševa kronično tretiranih tri-ortho-krezilfosfatom. Odgodena neuropatija uzrokovana organofosfatima mogla bi biti posljedicom prestanka djelovanja fosfolipaze NTE, što potom uzrokuje zatajenje endoplazmatskog retikuluma i smetnje u prijenosu signala putem aksona te interakcije između glija i aksona.

KLJUČNE RIJEČI: endoplazmatski retikulum, fosfolipid, karbamat, mozak, organofosfat

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