EXHALED BREATH CONDENSATE pH IN ADULT CROATIAN POPULATION WITHOUT RESPIRATORY DISORDERS: HOW HEALTHY A POPULATION SHOULD BE TO PROVIDE NORMATIVE DATA?

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The aim of this study was to obtain preliminary exhaled breath condensate (EBC) pH values for healthy adult Croatian subjects, and to evaluate criteria for defining respiratory health of population providing normal EBC pH values in epidemiologic studies. In 109 adults without a history of lower airway symptoms (AS), four groups were described by narrowing the definition of “health” down to 1) without lower AS; 2) without lower and upper AS; 3) without AS, with normal FEV1 and bronchial normoreactivity; 4) without AS, with normal FEV1, bronchial normoreactivity, normal total IgE, and with negative skin prick test. Median EBC pH values did not differ between the groups (7.72, 7.73, 7.73, 7.73), but as health criteria got stricter, we observed a slight, nonsignificant increase in minimal pH values (6.95, 7.10, 7.20, 7.37). Median EBC pH values with interquartile range in the total sample (7.72; 7.63 to 7.76) were within the range previously reported by other authors. They did not differ regarding sex, smoking habit and atopic status, and were not associated with age, FEV1 or total IgE. The non-significant trend in EBC pH observed with stricter criteria of respiratory health and atopic status indicates the need for further research on criteria for defining healthy population in a larger sample.

KEY WORDS: atopy, bronchial reactivity, EBC pH, health criteria, sex, skin prick test, smoking

As a completely non-invasive procedure, exhaled breath condensate (EBC) has been extensively studied in order to explore respiratory pathophysiology and its clinical relevance in the diagnosis and treatment of a variety of respiratory diseases, including asthma, chronic obstructive pulmonary disease, allergic rhinitis, pneumonia, adult respiratory distress syndrome, lung sarcoidosis, malignant lung tumours, cystic fibrosis, idiopathic lung fibrosis, and tuberculosis (1-3).

Many different volatile and non-volatile substances have been identified in EBC, such as carbon dioxide, ammonia, hydrogen peroxide, nucleotides, isoprostanes, leukotriens, nitric oxide, peptides, cytokines, and different ions (1, 4). There have also been attempts of biological monitoring of occupational exposure to substances, including metals and solvents (5-7). At present, none of these potential biomarkers have been sufficiently validated for clinical use (1, 4), and their application in larger-scale epidemiological studies is not very practical since the majority of them can be assessed in EBC only by expensive and technically demanding methods. An exception may be pH. It is considered to be the most validated parameter of EBC, which can be easily and reproducibly measured with non-expensive equipment (1). There are certain limitations and unresolved questions, including the source of airway acidification assessed by EBC pH, issues regarding the methodology of EBC collection, sample preparation and EBC pH measurement, as well as sensitivity and specificity.
Nevertheless, a number of studies have shown that acidification caused by the inflammation of the airways, like in asthma or chronic obstructive pulmonary disease, is reflected in lower EBC pH (8).

In order to implement this method in clinical practice and epidemiological studies, it is necessary to establish reference EBC pH values in a relevant population, as well as to validate the method in the laboratory. Presently, there are EBC pH values for more than 600 healthy adult subjects (1, 13). At the same time, criteria for defining “health” differ between studies, as shown in the Table 1. A great number of these subjects was selected as healthy, based on data obtained by a questionnaire alone or in combination with physical examination. In other studies, health criteria were rather strict, and included spirometry, non-specific bronchoprovocation test, and tests for objective atopy markers (total and specific serum IgE, skin prick test). This raises the question of which criteria are the most appropriate to define the health status for subjects who will provide normal, reference EBC pH values.

In this study we introduced a method for collecting EBC and measuring EBC pH in our laboratory, and we obtained preliminary EBC pH values for healthy, adult, smoking and non-smoking Croatian population. The other aim of the study was to see how EBC pH values vary with different criteria for defining respiratory health and atopic status in adult population.

SUBJECTS AND METHODS

Subjects and study protocol

The study involved 157 female and 43 male office workers from Zagreb, Croatia. All were volunteers, who signed an informed consent form. The subjects completed a questionnaire and underwent the following procedures: spirometry, non-specific bronchial challenge test, EBC collection, skin prick test with standard inhalatory allergens, and the analysis of total serum IgE level. Only the subjects who answered that they had never had lower airway symptoms (25 men, 84 women) were included in the study. In further analysis, the definition of “health” narrowed gradually, as described on Figure 1.

Additionally, 15 subjects (of whom five were men) reporting lower airway symptoms such as wheezing and/or dyspnoea with non-specific bronchial hyperreactivity and positive skin prick test to common inhalatory allergens, were enrolled as positive control.

The study was designed in accordance with the Declaration of Helsinki, and was approved by the Institute’s Ethics Committee.

Medical history

Using a simple questionnaire, we collected medical history data, including age, smoking habit, lower airway symptoms (including episodic dry cough not related to common cold, wheezing, chest tightness, and dyspnoea), and upper airway symptoms (including sneezing, rhinorrhea, nasal itching, and nasal obstruction not related to common cold). Smoking was analysed as a dichotomous variable (smoker or non-smoker).

Ventilatory function parameters

Forced expiratory volume in the first second (FEV₁) was determined using the standard method (32) with spirometer Pneumoscreen II (Jaeger, Würzburg, Germany). At least three measurements were recorded per subject, and the best value was used for analysis. FEV₁ was expressed and analysed as a percentage (FEV₁ %) of reference values (CECA II).

Nonspecific Bronchial Reactivity (NBR)

Nonspecific bronchial reactivity was assessed by means of a histamine challenge test, according to the procedure described by Chai et al. (33). The subjects inhaled doubling concentrations of histamine diphosphate solution (Sigma Chemical, St. Louis, MS) every three minutes from a DeVilbiss nebuliser (Model 646, DeVilbiss Health Care, Somerset, PA), controlled with a dosimeter (KoKo dosimeter, Ferraris Respiratory, Louisville, KY). The starting concentration of histamine diphosphate was 2 mg mL⁻¹, and the maximum dose used was 16 mg mL⁻¹. Bronchial responsiveness was measured by recording the subjects’ FEV₁ on a spirometer Pneumoscreen II (Jaeger, Germany) after each inhaled dose. Bronchial hyperreactivity was established if after the inhalation of ≤8 mg mL⁻¹ of histamine FEV₁ dropped ≥20 % of the value measured after the inhalation of the control solution, and further testing was stopped.

Total IgE

Total serum IgE antibodies were measured from venous blood samples using the enzyme-linked immunosorbent assay method (ELISA, IASON, Graz,
Table 1 Orally obtained EBC pH values in healthy adults

<table>
<thead>
<tr>
<th>Health criteria (reference)</th>
<th>Number of subjects</th>
<th>EBC sampling device (Deaeration with inert gas)</th>
<th>pH</th>
</tr>
</thead>
<tbody>
<tr>
<td>healthy, no details (14)</td>
<td>12</td>
<td>CB (no)</td>
<td>6.15±0.16</td>
</tr>
<tr>
<td>healthy non-smokers, no details (15)</td>
<td>30</td>
<td>ES (no)</td>
<td>6.30±0.30</td>
</tr>
<tr>
<td>healthy non-smokers, non-atopics, without respiratory symptoms, no history of lung disease; normal spirometry and NBR, negative SPT (16)</td>
<td>15</td>
<td>CB (no)</td>
<td>6.08 (5.58 to 6.64)</td>
</tr>
<tr>
<td>healthy, no details (spirometry was performed but not used as an exclusion criterion) (11)</td>
<td>10</td>
<td>CB (no)</td>
<td>7.24±0.24</td>
</tr>
<tr>
<td>healthy (general health based on questionnaire) (17)</td>
<td>21</td>
<td>RT (no)</td>
<td>6.17, 5.96 to 6.31</td>
</tr>
<tr>
<td>healthy non-smokers, normal spirometry (FEV₁) (18)</td>
<td>7</td>
<td>ES (no)</td>
<td>7.29±0.25</td>
</tr>
<tr>
<td>healthy non-smokers and smokers, no history of asthma, allergic rhinitis, hay fever, or atopic dermatitis (20)</td>
<td>270</td>
<td>RT (no)</td>
<td>8.09, 7.41 to 8.23</td>
</tr>
<tr>
<td>subjects without respiratory symptoms and history of asthma (21)</td>
<td>19</td>
<td>CB (argon)</td>
<td>7.65±0.20</td>
</tr>
<tr>
<td>healthy non-smokers, no history of allergy; normal spirometry and NBR, negative SPT (22)</td>
<td>10</td>
<td>CB (argon)</td>
<td>7.47±0.12, 7.49±0.10</td>
</tr>
<tr>
<td>healthy non-smokers, no details (23)</td>
<td>12</td>
<td>ES (argon)</td>
<td>7.46±0.48</td>
</tr>
<tr>
<td>healthy non-smokers, no history of significant chronic respiratory disease (24)</td>
<td>76</td>
<td>RT (argon)</td>
<td>7.70±0.49</td>
</tr>
<tr>
<td>subjects undergoing elective surgery, no history of chronic respiratory disease (25)</td>
<td>32</td>
<td>RT (argon)</td>
<td>7.90±0.23</td>
</tr>
<tr>
<td>healthy non-smokers, no details (25)</td>
<td>10</td>
<td>RT (argon)</td>
<td>7.90±0.30, 7.80±0.30</td>
</tr>
<tr>
<td>healthy, non-smokers, without respiratory tract infection within the last 4 weeks (26)</td>
<td>12</td>
<td>ES (argon)</td>
<td>7.61 (7.52 to 7.70)</td>
</tr>
<tr>
<td>healthy non-smokers, without pulmonary disease, non-atopics (negative SPT, not elevated total and specific IgE), normal spirometry, NBR and blood gas analysis (27)</td>
<td>15</td>
<td>ES (argon)</td>
<td>7.85±0.14</td>
</tr>
<tr>
<td>no history of lung disease, non-atopics (negative SPT), normal spirometry and NBR (28)</td>
<td>7</td>
<td>ES (argon)</td>
<td>7.90±0.10</td>
</tr>
<tr>
<td>healthy, without respiratory disorders, any acute or chronic systemic illness, and physician-diagnosed gastric disease (13)</td>
<td>404b</td>
<td>RT (argon)</td>
<td>8.00, 7.80 to 8.10 (4.50 to 8.40)</td>
</tr>
<tr>
<td>healthy non-smokers, without physician’s diagnosis of asthma, without respiratory symptoms; normal spirometry (FEV₁), negative bronchodilator test and SPT (29)</td>
<td>30</td>
<td>ES (argon)</td>
<td>7.54 (7.09 to 7.93)</td>
</tr>
<tr>
<td>healthy non-smokers with normal weight, without heart diseases, lung diseases and allergies (medical history and examination) (30)</td>
<td>15</td>
<td>RT (nitrogen)</td>
<td>8.20±0.13</td>
</tr>
<tr>
<td>healthy non-smokers, no history of respiratory disease; normal spirometry, normal NBR (31)</td>
<td>16</td>
<td>ES (argon)</td>
<td>6.72 (6.38 to 6.98)</td>
</tr>
</tbody>
</table>

*a mean ± SD; b median (range); c mean ± SEM; d median, interquartile range; e geometric mean ± SEM; f mean (95 % confidence interval); g range; h including 226 subjects older than 20 years; EBC – exhaled breath condensate; NAD – no abnormalities detected; NBR – non-specific bronchial reactivity; SPT – skin prick test; FEV₁ – forced expiratory volume in the 1st second; CB – custom built EBC sampling device; ES – EcoScreen; RT – RTube
Skin prick testing

Skin prick testing (SPT) was performed using a standard method (35) with a panel of common commercial inhalatory allergens: grass pollen mixture, birch, hazel, weed (*Ambrosia elatior*, *Artemisia vulgaris*) pollens, mites (*Dermatophagoidespteronyssinus*, *Dermatophagoides farinae*, and *Lepidoglyphus destructor*), cat, dog, and moulds (*Cladosporium herbarum* and *Alternaria alternata*). SPT included testing with positive control solution (10 mg mL\(^{-1}\) of histamine hydrochloride) and negative control solution (buffer solution). Skin reaction (wheal) was evaluated after 15 min. The mean skin reaction (mean wheal diameter) was calculated according to the formula \((D+d)/2\), where \(D\) represents the largest longitudinal diameter and \(d\) its midpoint orthogonal diameter in millimetres. For statistical evaluation, the difference between mean skin reaction to each allergen and negative control solution was used as a parameter of SPT reactivity. The results of SPT were considered positive (positive SPT) when the mean wheal diameter was larger than the negative control for more than 3 mm to at least one tested allergen.

Atopy status was defined as the presence of both elevated total IgE and positive SPT to at least one tested allergen.

**EBC collection and pH measurement**

All subjects were asked to fast for at least 12 h and refrain from smoking at least one hour before sampling. For the sampling, all subjects wore nose clips. Each subject provided a single EBC sample, breathing tidally into a commercial condenser (Eco Screen; Jaeger, Germany) for 15 min through a mouthpiece and a two-way non-rebreathing valve that also prevented saliva contamination due to integrated saliva trap. The condensate was collected into a Teflon-coated tube, which was disinfected and rinsed with tap water and wiped before sampling. After collection, the samples were frozen at -20 °C until analysis.

The samples were allowed to thaw to room temperature and were left exposed to ambient air until pH was stable (for three hours on average). pH was
measured using a Mettler pH meter (standard glass electrode-MP 220 Toledo, accuracy ±0.01) that had been calibrated with standard buffers (Mettler Toledo) at two points (pH 7.00 and pH 4.01).

Statistical analysis

The results are presented as mean values with standard deviations (age, FEV₁) or as medians with range (smoking index, total IgE, EBC pH) and interquartile range (EBC pH). Differences between groups (e.g. men and women, smokers and non-smokers, subject with and without upper airway symptoms) were tested with Student’s t-test (age, FEV₁), Mann-Whitney U test (smoking index, EBC pH, total IgE), or Fisher’s exact test (number of smokers, prevalence of respiratory symptoms, bronchial hyperreactivity, elevated IgE and positive SPT). Possible associations between EBC pH and age and FEV₁ and total IgE were analysed with Spearman’s correlation. Difference in EBC pH was tested between four overlapping groups defined as healthy according to criteria shown in Figure 1. The healthy groups were analysed as independent samples using the Kruskal-Wallis analysis of variance. EBC pH of each healthy group was also compared with positive control group using the Mann-Whitney U test. A P value of less than 0.05 was considered statistically significant in all analyses. Statistical analysis was performed using statistical software Stata/SE 10.0 for Windows (StatCorp LP, TX, USA).

RESULTS AND DISCUSSION

Table 2 shows the data about the age, smoking, respiratory parameters, and the prevalence of upper airway symptoms and positive objective atopy markers in subjects without a history of lower airway symptoms. The only difference found between men and women was a significantly higher prevalence of positive SPT and positive both objective atopy markers in men. The prevalence of positive SPT was more than two times higher, and the prevalence of positive both atopy markers seven times higher in men than in women. Only one female subject had FEV₁ lower than 80 % of the predicted value. Her FEV₁ was 78.5 %, and EBC pH 7.78.

Table 3 shows the profile of positive control subjects. Male to female ratio (approximately 1:3; Pearson χ²=0.78, P=0.355), the number of smokers (Pearson χ²=0.79, P=0.550), as well as age (t=1.502, P=0.136), did not differ significantly between subjects without lower airway symptoms and positive controls. Due to a very low number of smokers (three subjects) in the positive control group, smoking was not further analysed as a variable in this group. As expected, FEV₁ was lower in positive controls (t=-2.4673, P=0.015) and IgE levels were higher (z=4.082, P<0.0001) than in subjects without lower airway symptoms. In addition, the prevalence of upper airway symptoms was significantly higher in positive controls (14 out

### Table 2 General, respiratory, and atopic parameters in subjects without lower airway symptoms

<table>
<thead>
<tr>
<th>Subject (N)</th>
<th>Age / years Mean ± SD</th>
<th>Smoking status</th>
<th>Smoking index in smokers median, range</th>
<th>Upper airway symptoms N (%)</th>
<th>FEV₁ / % of expected value Mean ± SD</th>
<th>Bronchial hyperreactivity* N (%)</th>
<th>Total IgE</th>
<th>Positive SPT N (%)</th>
<th>Elevated IgE and positive SPT N (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total (109)</td>
<td>42.2 ± 9.0</td>
<td>34 (31.2)</td>
<td>200, 5 to 660</td>
<td>51 (46.8)</td>
<td>105.5 ± 11.8</td>
<td>13 (12.2)</td>
<td>24.3, 0 to 1000</td>
<td>16 (14.7)</td>
<td>32 (29.4)</td>
</tr>
<tr>
<td>Men (25)</td>
<td>43.4 ± 9.2</td>
<td>5 (20.0)</td>
<td>240, 10 to 600</td>
<td>9 (36.0)</td>
<td>106.3 ± 11.6</td>
<td>1 (4.0)</td>
<td>40.5, 0 to 000</td>
<td>7 (28.0)</td>
<td>13 (52.0)</td>
</tr>
<tr>
<td>Women (84)</td>
<td>41.8 ± 8.9</td>
<td>29 (34.5)</td>
<td>200, 5 to 660</td>
<td>42 (50.0)</td>
<td>105.2 ± 11.9</td>
<td>12 (14.6)</td>
<td>21.8, 0 to 424</td>
<td>9 (10.7)</td>
<td>19 (22.6)</td>
</tr>
</tbody>
</table>

Difference men vs women

<table>
<thead>
<tr>
<th>t</th>
<th>χ²</th>
<th>z</th>
<th>t</th>
<th>χ²</th>
<th>z</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.7655</td>
<td>1.894</td>
<td>0.658</td>
<td>1.517</td>
<td>0.389</td>
<td>1.501</td>
</tr>
<tr>
<td>0.446</td>
<td>0.222</td>
<td>0.510</td>
<td>0.258</td>
<td>0.698</td>
<td>0.292</td>
</tr>
</tbody>
</table>

* In 2 women NBR test was not performed due to contraindications; ' Serum sample from one subject was lost; † One woman had FEV₁ below 80 % (78.5 %); EBC – exhaled breath condensate; FEV₁ – forced expiratory volume in the 1st second; NBR – non-specific bronchial reactivity; SPT – skin prick test to standard inhalatory allergens; Differences between men and women were tested by Student’s t-test (age, FEV₁), Mann-Whitney U test (smoking index, total IgE), or by Fisher’s exact test (number of smokers, upper airway symptoms, bronchial hyperreactivity, elevated IgE, positive SPT, and elevated IgE + positive SPT). A P value of less than 0.05 was considered statistically significant.
of 15 subjects; Pearson $\chi^2$=11.45, Fisher’s exact $P=0.001$).

Table 4 shows median EBC pH values with interquartile range and minimum and maximum values for subjects without lower airway symptoms. They are within the range of values previously reported by other authors (Table 1). Table 4 also gives the values for positive controls.

In subjects without lower airway symptoms EBC pH was not associated with age (Spearman’s $r=0.049$, $P=0.611$), FEV$_1$, or IgE ($r=-0.148$, $P=0.126$). There were no differences in EBC pH between men (median, 7.73, interquartile range, 7.63 to 7.77) and women (7.71, 7.64 to 7.76; $z=0.126$; $P=0.900$), smokers (7.69, 7.48 to 7.77) and non-smokers (7.73, 7.65 to 7.75; $z=0.809$; $P=0.419$), subject without upper airway symptoms (7.73, 7.66 to 7.76) and subjects with upper airway symptoms (7.70, 7.53 to 7.76; $z=1.307$, $P=0.191$), subjects with bronchial normoreactivity (7.72, 7.63 to 7.76) and those with bronchial hyperreactivity (7.69, 7.64 to 7.75; $z=0.377$, $P=0.706$), and between atopics (7.76, 7.63 to 7.87) and non-atopics (7.71, 7.63 to 7.76; $z=1.221$; $P=0.222$). The correlations between EBC pH and age and FEV$_1$, and IgE were not significant in positive controls as well ($r=-0.153$, $P=0.586$; $r=0.248$, $P=0.372$; $r=-0.375$, $P=0.168$, respectively). Just like subjects without lower airway symptoms, positive control men and women did not differ in EBC pH ($z=-0.919$, $P=0.358$). It has repeatedly been shown by other authors that age, sex, and smoking have no effect on EBC pH (13, 17, 19, 20, 24). On the other hand, there are studies showing acute effects of smoking on certain EBC parameters, such as 8-isoprostane, H$_2$O$_2$, leukotriene B$_4$ and interleukin-6 levels (36-39), and EBC pH (40). The intensity of current smoking (number of cigarette packs smoked per day now) negatively correlated with EBC pH (40). However, it seems that active smoking does not affect EBC pH in subjects without a respiratory disease who refrain from smoking at least one hour before EBC collection (17, 24). The situation could be different in asthma patients, in whom smoking is associated with lower EBC pH than in non-smoking asthmatics (41). In our subjects with lower airway symptoms (positive control), however, the number of smokers was too low to allow statistical analysis, as mentioned above. To avoid possible acute effect of smoking on EBC pH, all subjects that entered the study did not smoke for at least one hour before EBC sampling.

EBC is believed to contain droplets of fluid lining the pulmonary surfaces (epithelial lining fluid), but their source may just as well be the upper respiratory tract and the upper gastrointestinal tract (42). In light of the concept of “united airways”, it is also possible that upper and lower airway disorders co-exist, and that the progression of atopic disease that manifests itself as allergic rhinitis can lead to acidification of the lower airways before asthma symptoms appear (43). This is supported by the findings that children with allergic rhinitis have lower EBC pH than healthy children, even in the absence of clinical signs of inflammation in the lower airways (43, 44). EBC pH in non-allergic upper respiratory disorders has not been studied so far. We did not observe an effect of the presence of upper airway symptoms on EBC pH in adult subjects without lower airway symptoms. Since these subjects were mainly non-atopics (99 out of 109), we could not establish a difference between the effects of atopic and non-atopic upper airway symptoms on EBC pH.

The lack of association between FEV$_1$, or non-specific bronchial reactivity and EBC pH in our study is no surprise as it enrolled only healthy subjects. A correlation between EBC pH and FEV$_1$, and IgE was observed in children with asthma (43) and in adult patients with asthma, chronic obstructive pulmonary disease (COPD), or bronchiectasis, but not in healthy control subjects (non-atopics with normal lung function and normal bronchial reactivity) (22). Data on EBC pH and non-specific bronchial reactivity are scarce. A positive trend (although not statistically significant) was found between EBC pH and non-specific bronchial reactivity in children with wheezing (45), but data for a population without airway symptoms are not available in literature.

As pointed out by Paget-Brown et al. (13) and Koutsokera et al. (1), there are plenty of studies of EBC pH which include healthy subjects, primarily to compare them with subjects with respiratory disease. Table 1 summarises the pH values of orally obtained EBC samples in healthy adults from different published studies. Between these studies, “health” criteria differ a lot. Differences are also substantial in the use of EBC sampling device, sampling procedure (e.g. duration), storage of samples (e.g. temperature and duration), sample preparation (native or treated with inert gas), and pH measurement (standard pH electrode, microelectrode, blood gas analyzer). An extensive study by Paget-Brown et al. (13), for example, included 404 healthy subjects of both sexes, with 226 subjects older than 20 years. The
### Table 3 General, respiratory, and atopic parameters in positive controls

<table>
<thead>
<tr>
<th>Subjects (N)</th>
<th>Age / year Mean ± SD</th>
<th>Smoking status</th>
<th>Upper airway symptoms Median, range</th>
<th>FEV1 / % of expected value Mean ± SD</th>
<th>Total IgE Median, range</th>
<th>Elevated N (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total (15)</td>
<td>46.0 ± 10.9</td>
<td>Smokers 3 (20.0)</td>
<td>75, 35 to 240</td>
<td>96.8 ± 18.8</td>
<td>180.6, 15.5 to 795.2</td>
<td>9 (60.0)</td>
</tr>
<tr>
<td>Men (5)</td>
<td>46.6 ± 15.3</td>
<td>Smoking index 1 (20.0)</td>
<td>35</td>
<td>89.8 ± 21.8</td>
<td>332.3, 180.6 to 795.2</td>
<td>5 (100.0)</td>
</tr>
<tr>
<td>Women (10)</td>
<td>45.7 ± 8.9</td>
<td>Median, range 2 (20.0)</td>
<td>158, 75 to 240</td>
<td>100.3 ± 17.4</td>
<td>74.4, 15.5 to 418.5</td>
<td>4 (40.0)</td>
</tr>
</tbody>
</table>

*Positive controls were subjects with lower airway symptoms, bronchial hyperreactivity, and positive skin prick test to common inhalatory allergens.

EBC – exhaled breath condensate; FEV1 – forced expiratory volume in the 1st second

### Table 4 EBC pH in subjects with different criteria for respiratory health and atopy, and in positive controls

<table>
<thead>
<tr>
<th>Subjects</th>
<th>N</th>
<th>pH Mean±SD</th>
<th>Median</th>
<th>Range</th>
<th>Interquartile range / P25-P75</th>
<th>Comparison with positive control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Without lower airway symptoms</td>
<td></td>
<td>7.67±0.21</td>
<td>7.72</td>
<td>6.95 to 8.26</td>
<td>7.63 to 7.76</td>
<td>z=-2.343, P=0.0191</td>
</tr>
<tr>
<td>Men</td>
<td>25</td>
<td>7.67±0.22</td>
<td>7.73</td>
<td>7.20 to 8.26</td>
<td>7.63 to 7.77</td>
<td></td>
</tr>
<tr>
<td>Women</td>
<td>84</td>
<td>7.67±0.20</td>
<td>7.71</td>
<td>6.95 to 8.10</td>
<td>7.64 to 7.76</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>109</td>
<td>7.67±0.21</td>
<td>7.72</td>
<td>6.95 to 8.26</td>
<td>7.63 to 7.76</td>
<td></td>
</tr>
</tbody>
</table>

Without lower and upper airway symptoms

Men | 16 | 7.71±0.15 | 7.75 | 7.20 to 7.87 | 7.68 to 7.77 |
Women | 42 | 7.68±0.17 | 7.73 | 7.10 to 7.96 | 7.66 to 7.75 |
Total | 58 | 7.69±0.16 | 7.73 | 7.10 to 7.96 | 7.66 to 7.76 | z=-2.756, P=0.0059 |

Without airway symptoms, with normal FEV1 and NBR

Men | 15 | 7.71±0.15 | 7.75 | 7.20 to 7.87 | 7.66 to 7.77 |
Women | 38 | 7.69±0.14 | 7.73 | 7.32 to 7.96 | 7.66 to 7.75 |
Total | 53 | 7.69±0.14 | 7.73 | 7.20 to 7.96 | 7.66 to 7.76 | z=-2.785, P=0.0054 |

Without airway symptoms, with normal FEV1, NBR and total IgE, and with negative SPT

Men | 7 | 7.71±0.06 | 7.70 | 7.64 to 7.78 | 7.66 to 7.77 |
Women | 32 | 7.70±0.14 | 7.73 | 7.37 to 7.96 | 7.67 to 7.77 |
Total | 39 | 7.70±0.13 | 7.73 | 7.37 to 7.96 | 7.66 to 7.77 | z=-2.771, P=0.0056 |

Positive controls

Men | 5 | 7.59±0.37 | 7.45 | 7.30 to 8.22 | 7.39 to 7.60 |
Women | 10 | 7.59±0.14 | 7.60 | 7.30 to 7.76 | 7.50 to 7.73 |
Total | 15 | 7.59±0.23 | 7.56 | 7.30 to 8.22 | 7.45 to 7.73 |

EBC – exhaled breath condensate; P25 – 25th percentile; P75 – 75th percentile; FEV1 – forced expiratory volume in the 1st second; NBR – non-specific bronchial reactivity; SPT – skin prick test to standard inhalatory allergens; Positive control – subjects reporting wheezing and/or dyspnoea with positive SPT and bronchial hyperreactivity. Median EBC pH values did not differ between the healthy groups (Kruskal-Wallis analysis of variance, P=0.817). The difference between healthy groups and positive control was tested by Mann-Whitney U test.
Figure 2 Box plots of EBC pH distribution in the four health categories and in positive control

0 – Positive control (subjects reporting wheezing and/or dyspnoea with positive SPT and bronchial hyperreactivity; N=15); 1 – Subjects without lower airway symptoms (N=109); 2 – Subjects without lower and upper airway symptoms (N=58); 3 – Subjects without airway symptoms, with normal FEV, and NBR (N=53); 4 – Subjects without airway symptoms, with normal FEV, NBR and total IgE, and with negative SPT (N=39); EBC – exhaled breath condensate; FEV – forced expiratory volume in the 1st second; NBR – non-specific bronchial reactivity; SPT – skin prick test to standard inhalatory allergens

Data are presented as medians with interquartile ranges, adjacent values, and minimal and maximal values.

health status was self-described by volunteers, and certain exclusion criteria were introduced to refine the selection. Excluded were smoking subjects, as well as those who reported chronic upper or lower airway symptoms, or symptoms related to viral respiratory tract infection (common cold). Similarly, the study of Vaughan et al. (24) on factors relevant to EBC pH monitoring involved 76 healthy non-smoking subjects whose respiratory health was defined as the absence of a “history of significant chronic respiratory disease”. The most recent study by Hauswirth et al. (20) included 270 healthy subjects of African ancestry whose health status was based on self-reported absence of a history of asthma, allergic rhinitis, hay fever, or atopic dermatitis. Studies in which respiratory and/or atopic status was more extensively evaluated usually involved a small number of subjects who served as a controls (16, 22, 27-29, 31). Which are the most appropriate health criteria for the selection of reliable and representative healthy control providing normal EBC pH values is still an unresolved issue that may be of particular interest for epidemiologic research. We defined our healthy subjects according to different criteria based on subjective and objective respiratory and atopic parameters. Four overlapping groups were described by narrowing the definition of “health” in four steps: in the 1st step we excluded subjects with lower airway symptoms (also the criteria for enrolment in the study); in the 2nd, from the group formed in the 1st step we excluded subjects with upper airway symptoms; in the 3rd, from the group formed in the 2nd step we excluded subjects with bronchial hyperreactivity and FEV, <80 %, and in the 4th step, from the group formed in previous step we excluded atopic subjects (with elevated IgE and positive SPT) (Figure 1, Table 4). Median values (7.72, 7.73, 7.73, and 7.73 for each respective step) and interquartile ranges were quite similar between the groups, and they showed no difference in EBC pH (Kruskal-Wallis ANOVA: $\chi^2=0.934$, $P=0.817$). However, the range of values narrowed with stricter criteria, and we observed a slight increase in minimal (6.95, 7.10, 7.20, and 7.37, respectively) and mean pH values (7.67, 7.69, 7.69, and 7.70, respectively) (Figure 2). This, albeit non-significant, trend indicates the need for further research on a larger sample. Since our sample size was limited, especially regarding healthy groups meeting stricter health criteria, any firm conclusion would be premature.

The pH values obtained from our participants are closer to values measured in samples after deaeration/decarbonation, although we did not treat the samples with inert gas (argon or nitrogen) to stabilise the pH. In general, literature describes three main approaches to EBC pH measurement: 1) in native samples, fresh or after defrosting; 2) after treating the samples with inert gas (bubbling or overlaying); and 3) at a standard CO level of the sample (1). The last method is proposed to be the most accurate (10). In our study, the samples were left at room temperature to permit gas exchange (primarily CO as a major EBC volatile component) with ambient air. We are aware that this procedure probably eliminates less CO from the sample then does deaeration/decarbonation with inert gas. Also, a greater variation in CO content (and consequently in pH value) is to be expected in our samples compared to deaerated/decarbonated samples, and especially to samples in which pH was measured at a standard CO level of the sample. Since our method has not yet been validated, in order to see whether it could discriminate between subjects with and without inflammatory changes in the airways, we introduced a positive control, i.e. subjects reporting lower airway symptoms typical for asthma (wheezing and/or dyspnoea) with positive SPT and bronchial hyperreactivity. We found that each healthy group had a significantly (0.16 to
0.17 units) higher EBC pH than positive control, indicating that the described method is able to detect airway acidification.

CONCLUSION

This study brings the first results on EBC pH in adult Croatian population without respiratory disorders. EBC pH values seem not to be affected by age, sex, smoking, upper airway symptoms, non-specific bronchial reactivity, or atopy. The established normal EBC pH range in our study is mildly alkaline and tight, and is comparable with data published in literature. Our data do not suggest that stricter health criteria for defining normal population bring an advantage for epidemiologic studies of EBC pH. Exclusion of subjects with respiratory symptoms and atopy did however show a slight trend toward more alkaline pH. This calls for further research in a larger number of subjects.

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Ciljevi preliminarnog istraživanja bili su izmjeriti pH-vrijednosti kondenzata izdaha (pH KI) odraslih stanovnika Hrvatske bez dišnih poremećaja te utvrditi kriterije potrebne za definiranje zdravlja dišnog sustava populacije u kojoj se planiraju utvrditi normativne vrijednosti KI-a. U uzorku od 109 odraslih osoba bez tegoba od strane donjih dišnih putova, sužavajući definiciju “zdravlja”, opisane su 4 skupine ispitnika: 1) bez donjih dišnih simptoma (DS); 2) bez gornjih i donjih DS; 3) bez DS i hiperreaktivnosti bronha s normalnim FEV₁; 4) bez DS i hiperreaktivnosti bronha s normalnim FEV₁, ukupnim IgE i s negativnim prick testom. Medijani pH-vrijednosti nisu se razlikovali između skupina (7,72; 7,73; 7,73; 7,73), ali uvođenjem sve strožijih kriterija zdravlja uočen je blag, iako nesignifikantan, porast minimalnih pH-vrijednosti KI-a (6,95; 7,10; 7,20; 7,37). Medijan pH KI s interkvartilnim rasponom u ukupnom uzorku (7,72; 7,63 do 7,76) bio je unutar raspona vrijednosti izmerenih u istraživanjima drugih autora. Na pH KI nisu utjecali spol, navika pušenja i atopijski status i nije bio povezan s dobi, vrijednostima FEV₁ ili ukupnim IgE. Uočeni nesignifikantni trend porasta pH KI nakon uvođenja strožijih zdravstvenih kriterija sugerira potrebu daljnjih istraživanja kriterija za definiranje zdravlja dišnog sustava na većem uzorku.

KLJUČNE RIJEČI: atopija, kožni prick test, kriteriji zdravlja, pH KI, pušenje, reaktivnost bronha, spol

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