Evaluation of amphetamine-type stimulant abuse through hair analysis: Results from 12 years of work

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Received in March 2014
CrossChecked in March 2014
Accepted in June 2014

Hair analysis is a reliable tool for detecting long-term exposure to illegal drugs, including amphetamine-type stimulants, over periods from a few weeks to a few months, depending on the length of the hair used for analysis. Between 2000 and 2012, over 600 hair samples were analysed at the Institute for Medical Research and Occupational Health, Croatia (IMROH) for the presence of amphetamine-type stimulants. IMROH has used the same procedure for testing hair samples for amphetamine-type stimulants for over twelve years. It was found to be reliable for confirming repeated abuse of amphetamine-type stimulants. Gas chromatography/mass spectrometry (GC/MS) was used to determine amphetamine, methamphetamine, 3,4-methylenedioxyamphetamine (MDA), 3,4-methylenedioxymethamphetamine (MDMA-Ecstasy), and 3,4-methylenedioxyethylamphetamine (MDEA) in hair. Hair samples were either taken at the Institute, delivered by mail or a third person brought them to the laboratory. In most cases, the hair samples were tested anonymously. A total of 23 % of the tested samples were positive for one or more amphetamine-type stimulant. MDMA was the most frequently detected substance, whereas the most frequent combination was amphetamine with MDMA. Our results could indicate a trend in amphetamine-type stimulant abuse among young people in the Republic of Croatia.

KEY WORDS: GC/MS; MDMA; methamphetamine; quantitative determination
stimulants seized, a rise was recorded in 2011 (4). In a comprehensive study performed in a Zagreb wastewater treatment plant in 2009, a consumption of 1–3 kg per year amphetamine-type stimulants was estimated based on their analysis in wastewater. The authors concluded that the amphetamine-type drug consumption rate in the city of Zagreb (3.6–9.7 mg per day in 1000 inhabitants) is comparable to that in Italy and Switzerland, but significantly lower than in UK and Spain (5).

Due to the abovementioned reasons, the early identification of drug abuse in the general population is critical in planning preventative action and reducing the damage caused by illicit drug use. Unlike urine, in which the generally accepted detection time for amphetamine-type stimulants is 1 to 3 days, hair provides information on long-term exposure to amphetamine-type stimulants over a period from a few weeks to a few months, depending on the length of the hair collected (6).

Since 2000, the Institute for Medical Research and Occupational Health in Zagreb, Croatia (IMROH) has provided quantitative hair analyses for amphetamine-type stimulants for a wide variety of customers including private customers, addiction prevention centres, hospitals, courts, and employers. The aim of this paper is to provide an overview of amphetamine-type stimulant abuse in Croatia, as well as to evaluate the different characteristics of the hair samples analysed at the Institute with regard to how and from whom they were received.

MATERIALS AND METHODS

Hair samples

Hair samples (n=666) from various sources were received or taken in the laboratory. A strand of hair of approximately 5 mm in diameter was cut from close to the scalp at the vertex posterior area, folded in aluminium foil or paper, and the proximal and distal ends were marked. We analysed hair segments 2-4 cm long from the proximal end, which represent approximately 2-4 months of hair growth as stated in the literature (7). The study was conducted according to the ethical standards of the Helsinki Declaration and was approved by the IMROH Ethics Committee. The investigation presented in this paper was performed within a wider research project entitled “Identification of drug abuse by comparative analysis of biological specimens” and funded by the Ministry of Science and Technology of the Republic of Croatia.

Hair analysis

Hair samples were analysed according to a previously validated procedure (8). The gas chromatography/mass spectrometry (GC/MS) method was developed for determining amphetamine, methamphetamine, MDMA, MDA, and MDEA in hair. Over the years, a few procedural modifications were done (reagent and standard suppliers were changed and deuterated internal standards were purchased). All of the changes were fully validated before routine laboratory work. In brief, the hair was washed in dichloromethane, dried, cut into very small pieces, and 50 mg of hair was used for analysis. Hair samples were submitted to alkaline digestion with 1 mol L⁻¹ sodium hydroxide at 70 °C for 20 min prior to extraction with ethyl acetate (2x1 mL). The extract was evaporated to dryness in the presence of a 100 μL mixture of methanol:hydrochloric acid (99:1, v/v) to prevent a loss of volatile amphetamine and derivatized with 50 μL of heptafluorobutyric anhydride (HFBA) at 60 °C for 30 min. The derivatized samples were evaporated to dryness, and reconstituted in 100 μL of ethyl acetate for injection of 1 μL onto a gas chromatograph Varian 3400 CX with Saturn ion trap mass spectrometer equipped with HP-5MS capillary column (5 % diphenyl-95 % dimethylpolysiloxane, 30 m, 0.25 mm ID, 0.25 μm film thickness; J&W Agilent Technologies; Santa Clara, USA). Three ions for the analytes and two ions for the internal standards were monitored in the selected ion monitoring (SIM) mode. The following ions were used for each drug: amphetamine-HFBA, m/z 240, 118, 91; methamphetamine-HFBA, m/z 254, 210, 118; MDA-HFBA, m/z 135, 162, 240; MDMA-HFBA, m/z 268, 135, 162; amphetamine-d₅-HFBA, m/z 244, 92; and MDMA-d₅-HFBA, m/z 258, 165. The underlined ions were used for quantitation. The standards were prepared in blank hair extracts. Each case sample batch included standards and positive and negative control. The precision expressed as relative standard deviation (RSD) was <9 % and the accuracy was >85 % for all of the 5 analytes. The limit of detection ranged from 0.01 ng mg⁻¹ to 0.05 ng mg⁻¹. The confirmation cut-off for amphetamine-type stimulants was 0.2 ng mg⁻¹. External quality assessment was performed through participation in the Proficiency Testing programme.
organised by the Society of Hair Testing (9). For the purpose of this study, all data above the limit of detection were included.

RESULTS AND DISCUSSION

During the period from 28 April 2000 to 10 December 2012, 666 hair samples were analysed. A large majority of our customers were parents who had suspected their children of taking drugs – primarily amphetamine-type stimulants. In most instances, a third person (e.g., parent) brought the hair sample for analysis (40 %). In other cases, hair samples were taken at IMROH by trained personnel (37 %) or delivered by mail (23 %). In 73 % of cases, hair samples were tested anonymously (e.g., the hair samples were received under only the first or a fictitious name, or a number). Hair analysis requests from private customers accounted for approximately 92 % of the total number of samples. Less than 8 % of cases included hair samples tested for official purposes [addiction prevention centres and hospital requests (3.8 %), workplace drug testing (2.7 %), and court proceedings (1.4 %)]. The probable reason for such a low percentage of official requests in Croatia compared to e.g. United Kingdom (10) is that hair testing for illegal drugs has not yet been included in any Croatian law or regulation.

The main drawbacks in case of sampling hair from a third person are the lack of control regarding protocols and the ability to collect hair "next to the scalp". LeBeau et al. (11) suggested that, beside the variability in the growth rate, inconsistent collection may have an important impact on the interpretation of results.

Among customers who could be identified by gender (they came personally to the laboratory) males were more likely to be positive (23 % of positive males vs. 17 % of positive females), which is in accordance with studies by Strote et al. (12), Martins et al. (13), and Molinaro et al. (14), who reported a higher prevalence of illegal drug use among males. Also, males were 2.5 times more frequent clients compared to females.

Table 1 summarises the positive results of hair testing (n=155), in which one or more type of drug were found. A total of 23 % of the tested samples were positive for one or more amphetamine-type stimulant. Among the hair samples received, 25 % of samples from users who provided their identity tested positive compared to 20 % of positive hair samples received anonymously. Considering single drug use, MDMA was the most frequent detected substance (in 10 % of the samples), followed by amphetamine (5 %). The most frequent combination was amphetamine with MDMA (5 % of all hair samples). Methamphetamine was detected in only 1 % of hair samples. In less than 2 % of hair samples, we detected methamphetamine + MDMA, amphetamine + methamphetamine, amphetamine + methamphetamine + MDMA or amphetamine + MDMA + MDA + MDEA.

According to surveys conducted on a large sample size in Norway (15), Australia (16), United States (13),
and Taiwan (17), MDMA appeared to be the most commonly used illegal drug in adolescents and young adults. The prevalence of MDMA in our study was not surprising since most of our clients were adolescents and young adults considered to be frequent consumers of MDMA. In addition, MDMA is easily available and relatively cheap. Growing concerns have been raised about ecstasy use during adolescence because young ecstasy users tend to be more naive and more vulnerable to the harmful effects. The small number of samples positive for methamphetamine could be explained by its low accessibility and relatively high price (4). The levels detected for amphetamine-type stimulants are presented in Table 2.

The measured concentrations were in accordance with already published concentration ranges (10, 18, 19). Among the 109 cases positive for MDMA, its metabolite MDA was detected in 23 cases. The MDA/MDMA ratio in hair ranged from 0.01 to 0.25, which was in agreement with earlier observations (20). In most of the MDMA positive samples (73.4 %), MDMA was found in mass fractions up to 5.0 ng mg⁻¹, which is in accordance with previously reported results (21). Amphetamine was found in 77 cases, 36 of which exhibited a co-occurrence with MDMA, which is generally the most frequent combination. The probable reasons for the wide range of mass fractions for amphetamine and MDMA could be the different purity, frequency, and amount consumed. Also, the different hair treatments in 10 % of the analysed hair samples could be the reason for a slight decrease of drug content in those hair samples, which has been documented by several authors (18, 22, 23).

Interpretation of hair analysis results is a very challenging process due to several possible sources of error. In addition to different hair growth rate and errors in sampling, a possible problem in hair analysis could be false positive hair testing result caused by passive exposure to drug from the environment. Therefore, a decontamination step using appropriate solvents is necessary prior to the analysis of hair samples (24). In addition, another problem is the lack of correlation between frequency of drug use and its concentration in hair due to the different incorporation rate of drug in the hair, different hair melanin content, use of hair treatments (bleaching and dyeing), etc. Pharmacogenetic variations also play an important role in the interpretation of hair analysis results (25).

**CONCLUSIONS**

It is evident that hair is a reliable biological marker for cumulative exposure to illicit drugs. IMROH has used the same hair sample testing procedure for amphetamine-type stimulants for over twelve years. This method was found to be reliable for confirming repeated amphetamine-type stimulant abuse. Our study can serve to provide a preliminary idea about the trend of amphetamine-type stimulant abuse among adolescents and young adults in Croatia.

**Acknowledgements**

Author’s attendance of the TIAFT2013 Congress (where this study was presented) was supported by a donation from Biovit, Croatia. The authors also wish to thank Ms Vesna Triva for her most skillful technical assistance.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Number of positive samples</th>
<th>Mass fraction (ng mg⁻¹) median (range)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MDMA</td>
<td>109</td>
<td>2.30 (0.45-90.8)</td>
</tr>
<tr>
<td>Amphetamine</td>
<td>77</td>
<td>1.51 (0.15-118.7)</td>
</tr>
<tr>
<td>MDA</td>
<td>23</td>
<td>0.81 (0.18-4.64)</td>
</tr>
<tr>
<td>Methamphetamine</td>
<td>20</td>
<td>0.67 (0.10-2.01)</td>
</tr>
<tr>
<td>MDEA</td>
<td>1</td>
<td>2.13 (-)</td>
</tr>
</tbody>
</table>

MDMA - 3,4-methylenedioxyamphetamine; MDA - 3,4-methylenedioxyamphetamine; MDEA - 3,4-methylenedioxyethylamphetamine
REFERENCES


Sažetak

Evaluacija zlouporabe stimulansa amfetaminskog tipa putem analize kose: rezultati dvanaestogodišnjeg rada

Analiza kose pouzdan je način za detektiranje dugotrajnog konzumiranja ilegalnih droga, uključujući stimulanse amfetaminskog tipa u razdoblju od nekoliko tjedana do nekoliko mjeseci, ovisno o duljini kose koja se koristi za analizu. Od 2000. do 2012. u Institutu za medicinska istraživanja i medicinu rada analizirano je više od 600 uzoraka kose na prisutnost stimulansa amfetaminskog tipa. Vezani sustav plinski kromatograf/spektrometar masa koristio se za određivanje masenog udjela amfetamina, metamfetamina, 3,4-metilenedioksiamfetamina (MDA), 3,4-metilenedioksimetamfetamina (MDMA-Ekstazi) i 3,4-metilenedioksietilamfetamina (MDEA) u kosi. Uzorci kose uzimani su u Institutu, dostavljeni su poštom ili ih je treća osoba donijela u laboratorij. U većini slučajeva uzorkovanje i analiza kose obavljeni su anonimno. Dvadeset tri posto analiziranih uzoraka kose bilo je pozitivno na jedan ili više stimulansa amfetaminskog tipa. Najčešće je detektiran MDMA, a najčešća kombinacija stimulansa bila je amfetamin s MDMA. U Institutu se postupak za testiranje uzoraka kose na prisutnost stimulansa amfetaminskog tipa provodi više od 12 godina i smatra se pouzdanim načinom potvrde ponovljenog konzumiranja tih stimulansa. Naši rezultati mogu upućivati na trend zloporabe stimulansa amfetaminskog tipa među mlađom hrvatskom populacijom.

KLJUČNE RIJEČI: GC/MS; kvantitativno određivanje; MDMA; metamfetamin

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