ABSTRACT

The renin-angiotensin system (RAS) has been implicated in the pathogenesis of acute and chronic pancreatitis. Angiotensin-converting enzyme (ACE) is the key enzyme which activates RAS. The ACE intron 16 insertion/deletion (I/D) polymorphism is associated with ACE activity and is considered to be a risk factor for several inflammatory processes. We investigated this polymorphism in 68 patients with acute pancreatitis (AP) and 157 healthy Turkish control subjects. Patients were evaluated with ultrasonography, abdominal tomography and laboratory markers and grouped by status for diabetes mellitus (DM), hypertension (HT), and both these diseases and by etiology. Genotyping of the I/D polymorphism was performed by polymerase chain reaction (PCR). The DD genotype was more prevalent in healthy controls, however, genotype II was significantly more frequent in AP patients (p<0.05). In severe AP patients, the genotype II frequency was significantly higher than in controls (p<0.05). Acute pancreatitis patients with both DM and HT had lower frequencies of genotype DD and of the D allele, and higher frequencies of genotype II and of the I allele than patients with either DM or HT (p<0.05).

Key words: Acute pancreatitis (AP), Angiotensin-converting enzyme (ACE) gene polymorphism, Genetic polymorphism

INTRODUCTION

Acute pancreatitis (AP) is an acute inflammatory process of variable of etiology [1]. While etiology and pathogenesis of AP have been intensively investigated, it is still unclear why some patients progress to organ failure and others do not [2,3]. Blockage of the duodenal papilla or ampulla of Vater is the common characteristic of the disease. In developed countries, obstruction of the common bile duct by stones (38%) and alcohol abuse (36%) are the most frequent causes [4,5], but iatrogenic factors, sphincter of Oddi dysfunction and eating disorders are also important [6]. Acute pancreatitis is viewed as an event and chronic pancreatitis as a process that is sequentially linked to AP and reflects a complex interaction between genetic and environmental factors [7]. The major common genetic risk factors have yet to be defined [8].

The renin-angiotensin system (RAS) has been investigated in the pathogenesis of several diseases. Local RAS components exist in brain, heart, kidney, pancreas, adrenal glands and gonads [9-11], and contribute in the regulation of cell growth, differentiation, proliferation and apoptosis, reactive oxygen species generation, tissue inflammation and fibrosis, and hormonal secretion [12]. Renin-angiotensin system has been implicated in pathogenesis.
ACE GENOTYPE IN AP IN TURKEY

of AP and chronic pancreatitis [8]. Although the etiology of AP is believed to be multifactorial, the activation of proteolytic enzymes, lipase, kinins and other active peptides may be responsible for alterations of RAS expression [13,14]. In fact, the activity of the plasma RAS is significantly increased in AP [15,16]. Renin-angiotensin system is important in regulation of electrolyte balance, fluid and blood pressure. Angiotensin-converting enzyme is the key enzyme which activates the RAS [8,9] by converting angiotensin I to angiotensin II, which is a potent vasoconstrictor. The ACE inactivates bradykinin, a vasodilator of the kallikrein-kinin system, which has major influence in inflammatory processes. Since angiotensinogen and angiotensin receptors may play a role in induction of inflammation and microcirculatory regulation in the pancreas, they may contribute to its injury in AP [16,17]. Association of severe AP and impairment of pancreatic microcirculation has been demonstrated in experimental models of AP [18]. Indeed, vasoconstriction, capillary stasis, decreased oxygen tension, and progressive ischemia occur early in the course of AP [19].

The ACE gene is located on human chromosome 17q23. Three genotypes are associated with an Alu repetitive sequence about 287 bp long on intron 16. These genotypes are insertion (I) and deletion (D) alleles, respectively [16,17].

The DD genotype has been linked to several inflammatory diseases [8,9] and results in higher levels of circulating ACE than the II and DI genotypes. It is also significantly more frequent in patients with myocardial infarction or diabetic proteinuria than in controls [10]. However, several studies have reported that the ACE I/D polymorphism was not a risk factor in AP and chronic pancreatitis [8,9]. In this study, we investigated the I/D polymorphism in AP.

MATERIALS AND METHODS

Sixty-eight patients with AP and 157 healthy controls were recruited from the archives of the Department of Gastroenterology, Ege University Medical School, Izmir, Turkey. Informed written consent was obtained from each patient and the study protocol conforms to the ethical guidelines of the declaration of Helsinki. Etiology, gender, age, clinical presentation, clinical course, genotype frequency of the ACE I/D polymorphism and their association were compared.

Blood specimens from all participants were obtained with a standard venipuncture technique that used ethylenediamine tetra-acetic acid-containing tubes. DNA was isolated from peripheral blood by a standard phenol/chloroform extraction method [12].

For the ACE polymorphism, we used polymerase chain reaction (PCR) methodology [19], with the upstream primer 5’-CTG GAG ACC ACT CCC ATC CTT TCT-3’ and the downstream primer 5’-GAT GTG GCC ATC ACA TTC GTC AGA T-3’.

Amplification was performed for 35 cycles with denaturation, extension and annealing temperatures of 94.8°C, 60.8°C and 72.8°C, respectively. The resulting PCR products were separated on 2% agarose gel with ethidium bromide staining and were visualized under ultraviolet light. Homozygotes for the deletion or insertion genotypes were described as DD and II, respectively, and the heterozygous genotype as ID.

All statistical analyses were performed using the SPSS 11.0 statistical program. Genotypes, allele frequencies, clinical features at diagnosis were evaluated by the χ² test. Clinical data are reported as mean ± SD (standard deviation) and as percentages. Statistical significance was taken as p <0.05.

RESULTS

The characteristics of the patients with AP and the healthy controls are summarized in Table 1. Etiology and severity of AP are summarized in Table 2. The etiology was classed as biliary in 44, and 24 were classed to be of other etiologies [six hypertriglyceridermia, 10 post endoscopic retrograde cholangiopancreatography procedure (ERCP), three alcoholic, five idiopathic]. Fifty-five patients were diagnosed to have mild AP, 13 patients were diagnosed with severe AP. Thirteen patients with AP had diabetes mellitus (DM) type 2, 23 had essential hypertension (HT), and 10 had both.

Genotype distribution of ACE gene for both groups is summarized in Table 3. The DD genotype was more prevalent among healthy controls than in AP patients, while the II genotype was more prevalent in AP patients than healthy controls (p <0.05).

Genotype distribution of the ACE gene in bil-
Table 1. Characteristics of the study groups

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Acute Pancreatitis</th>
<th>Healthy Controls</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of subjects</td>
<td>68</td>
<td>157</td>
</tr>
<tr>
<td>Sex (males/females)</td>
<td>42/26</td>
<td>102/55</td>
</tr>
<tr>
<td>Age (years)</td>
<td>$51 \pm 5$</td>
<td>$24 \pm 7$</td>
</tr>
<tr>
<td>Diabetes mellitus</td>
<td>13</td>
<td>0</td>
</tr>
<tr>
<td>Hypertension</td>
<td>23</td>
<td>0</td>
</tr>
<tr>
<td>Diabetes mellitus + hypertension</td>
<td>10</td>
<td>0</td>
</tr>
</tbody>
</table>

Table 2. Etiology and severity of acute pancreatitis

<table>
<thead>
<tr>
<th>Etiology</th>
<th>Mild AP ($n$)</th>
<th>Severe AP ($n$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Biliary</td>
<td>34</td>
<td>10</td>
</tr>
<tr>
<td>Endoscopic retrograde cholangiopancreatography procedure (ERCP)</td>
<td>9</td>
<td>1</td>
</tr>
<tr>
<td>Hypertriglyceridemia</td>
<td>4</td>
<td>2</td>
</tr>
<tr>
<td>Alcohol</td>
<td>3</td>
<td>0</td>
</tr>
<tr>
<td>Idiopathic</td>
<td>5</td>
<td>0</td>
</tr>
<tr>
<td>Severity</td>
<td>55</td>
<td>13</td>
</tr>
<tr>
<td>Ranson Score (median range)</td>
<td>1 (0-4)</td>
<td>4 (1-8)</td>
</tr>
</tbody>
</table>

Table 3. Genotype distribution of the angiotensin-converting enzyme gene in the study groups

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Acute Pancreatitis $n$ (%)</th>
<th>Healthy Controls $n$ (%)</th>
<th>$p$ Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>DD</td>
<td>17 (25.0)</td>
<td>61 (38.0)</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>DI</td>
<td>37 (54.4)</td>
<td>82 (52.0)</td>
<td>N.S.</td>
</tr>
<tr>
<td>II</td>
<td>14 (20.0)</td>
<td>14 (8.0)</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>D (allele frequency)</td>
<td>71 (52.2)</td>
<td>204 (64.9)</td>
<td>N.S.</td>
</tr>
<tr>
<td>I (allele frequency)</td>
<td>65 (47.8)</td>
<td>110 (35.1)</td>
<td>N.S.</td>
</tr>
</tbody>
</table>

N.S.: not significant.

There was no significant difference in the distribution of the I/D polymorphism or in allele frequency between biliary and non biliary AP.

Genotype distribution of the ACE gene in AP associated with DM and/or HT is summarized in Table 5. There is no significant difference in the distribution of the I/D polymorphism or allele frequency between AP with DM and AP with HT.

However in patients with both these diseases, the ACE II genotype and allele I frequency was significantly higher than in the healthy group or in patients with only one of these diseases ($p < 0.05$). The distribution of ACE genotypes DD and DI were similar in patients with mild or severe AP. However, frequency of the II genotype in severe AP was significantly higher than healthy controls ($p < 0.05$) (Table 6).
DISCUSSION

The subjects who had the homozygous deletion (DD) genotype had the highest ACE frequency, while subjects who had the homozygous insertion (II) genotype had the lowest ACE levels. Studies from the United States, German and Finnish populations did not find a relationship between AP and I/D polymorphisms [9,20]. We found the DD genotype to be more prevalent in healthy controls than in patients with AP, however, the II genotype was significantly more prevalent in AP patients than in healthy controls ($p<0.05$). Even so, the ACE II genotype was significantly higher in severe AP pa-
patients than in healthy controls, and ACE II was not related to the etiology of pancreatitis.

Studies in the European populations, revealed the ratio for the ACE genotype DD/ID/II to be 1:2:1. However, in Turkey, the DD genotype is the most frequent, and genotype II the least common in the healthy population [21-23]. This may be one of the reasons for the difference between our results and those from studies of the United States, German and Finish populations [9,20]. A relationship between the II genotype and inflammation and pancreatic injury needs further clarification. It has been suggested that the DD genotype is related to essential HT and DM [23,24]. On the other hand, the association between the I/D polymorphism and HT and DM is still controversial [24-27]. A study from Turkey did not show a significant association between the ACE gene polymorphism and HT in diabetic patients [26]. A study from Sweden showed that the DD genotype increases the risk of HT in diabetic patients [28].

We did not find significant difference either in the distribution of the I/D genotype polymorphism or allele frequency in diabetic AP and hypertensive AP groups. We have found that the ACE II genotype and allele I frequencies are higher in the patients who have both diabetes and HT compared to patients who have only one of these diseases. In conclusion, the ACE gene polymorphism may play a role in AP. We need larger studies for investigating the relation of the ACE I/I genotype with other inflammatory parameters. By such studies we could evaluate the significance of the ACE I/I genotype in AP and its possible use as a genetic marker.

REFERENCES


